



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 : C07K 16/28, 14/435, C12N 15/00, 15/11, 15/12, 5/10, G01N 33/53, A61K 39/395	A1	(11) International Publication Number: WO 97/42224 (43) International Publication Date: 13 November 1997 (13.11.97)
--	----	---

(21) International Application Number: PCT/US97/07737

(22) International Filing Date: 6 May 1997 (06.05.97)

(30) Priority Data:

08/643,798

7 May 1996 (07.05.96)

US

08/851,160

5 May 1997 (05.05.97)

US

(71) Applicant: MILLENNIUM PHARMACEUTICALS, INC.
[US/US]; 640 Memorial Drive, Cambridge, MA 02139-
4815 (US).(72) Inventor: PAN, Yang; 5 Eliot Street, Brookline, MA 02167
(US).(74) Agent: FASSE, J., Peter; Fish & Richardson, P.C., 225 Franklin
Street, Boston, MA 02110-2804 (US).(81) Designated States: AU, CA, JP, European patent (AT, BE,
CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL,
PT, SE).

Published

With international search report.

(54) Title: NEUROTACTIN AND USES THEREFOR

(57) Abstract

The present invention relates to the identification and characterization of a novel, membrane-anchored chemokine, neurotactin. Sequence analysis of neurotactin reveals that, while it includes an amino terminal domain which resembles that of other chemokines, it has an overall structure which distinguishes it from all presently identified chemokines. Neurotactin is highly expressed in normal mammalian brain.

Best Available Copy

Best Available Copy

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

- 1 -

NEUROTACTIN AND USES THEREFORBackground of the Invention

5 This invention relates to a new chemokine, neurotactin, and methods of preparing and using neurotactin.

Chemokines are proteins involved in the activation and chemotaxis of leukocytes. They are believed to be
10 important mediators of inflammation (Baggiolini et al., *Immunology Today* 15:127, 1994).

Chemokines have been divided into three families. In chemokines of the C-X-C family, one amino acid separates the first two cysteines. Chemokines in this
15 family are thought to be involved in the chemotaxis of neutrophils, induction of changes in cell shape, transient increase of intracellular calcium, granule exocytosis, and respiratory burst. Interleukin-8 (IL-8), neutrophil activating protein-2 (NAP-2) and granulocyte
20 chemotactic protein (GCP) belong to this class. All known C-X-C chemokines have been mapped to human chromosome 4 and mouse chromosome 5.

In the C-C family, the first two cysteines are adjacent to one another. Members of this family are
25 chemotactic for monocytes, but not neutrophils. Recent studies have shown that they are capable of activating basophils and eosinophils. Proteins belonging to the C-C class of chemokines include monocyte chemotactic proteins 1, 2, and 3 (MCP-1, MCP-2, and MCP-3), RANTES,
30 and macrophage inflammatory proteins α and β (MIP-1 α and MIP-1 β). Recently, MIP-3, MIP-4, and MIP-1 γ have also been described (WO 95/17092). All known C-C chemokines have been mapped to human chromosome 17 and mouse chromosome 11.

- 2 -

An example of a third class of chemokine has also been identified. This chemokine, lymphotactin, was isolated from progenitor T lymphocytes. Lymphotactin is chemotactic to lymphocytes (Kelner et al., *Science* 5 266:1395, 1994). Unlike the chemokines of the C-C and C-X-C families in which two disulfide bonds stabilize the protein, lymphotactin has only one disulfide bond. Lymphotactin was mapped to human and mouse chromosome 1.

A variety of cell types are involved in the 10 various inflammatory states. For example, acute infiltrates found after bacterial infection are mainly neutrophilic, while mononuclear cells predominate after infection by an intracellular pathogen. Basophils and eosinophils dominate in both immediate-type allergic 15 response and autoimmune diseases. Increased understanding of the regulation of these various cell types by chemokines will facilitate the development of more effective therapies for disorders related to inflammation.

20 Brain inflammation is only partially understood. It appears that the brain regulates its own immune response rather than being an immunological privileged organ (Trevor et al., *Immunology Today* 15:566, 1994). Inflammatory cytokine production in the brain is 25 initiated by infiltrating T cells but longer term inflammation is dependent on CNS resident cells, such as microglial cells.

Summary of the Invention

The present invention relates to the 30 identification and characterization of neurotactin, a novel membrane-anchored chemokine. Sequence analysis of neurotactin reveals that, while it includes an amino terminal domain which resembles that of other chemokines, it has an overall structure which distinguishes it from 35 all presently identified chemokines.

- 3 -

The expression pattern of neurotactin is unusual for a chemokine in that it is highly expressed in normal mammalian brain. Similar to other chemokines, neurotactin is upregulated in bone marrow stromal cells, endothelial cells, and fibroblasts which have been treated with lipopolysaccharide (LPS) and phorbol myristate acid (PMA), both of which are activators.

A murine form of neurotactin described herein encodes a protein of 395 amino acids (Fig. 1). This murine form of the protein (SEQ ID NO:2) begins with an approximately 21 amino acid long signal sequence followed by an apparent extracellular domain of approximately 318 amino acids extending from about amino acid 22 to about amino acid 339; a transmembrane domain of approximately 21 amino acids extending from about amino acid 340 to about amino acid 360; and a cytoplasmic domain of approximately 35 amino acids extending from about amino acid 361 to amino acid 395.

A human form of neurotactin described herein encodes a protein of 397 amino acids (Fig. 2). This human form of the protein (SEQ ID NO:4) begins with an approximately 21 amino acid long signal sequence followed by an apparent extracellular domain of approximately 321 amino acids extending from about amino acid 22 to about amino acid 341; a transmembrane domain of approximately 22 amino acids extending from about amino acid 342 to about amino acid 362; and a cytoplasmic domain of approximately 35 amino acids extending from about amino acid 363 to amino acid 397.

Within the extracellular domain of both the murine form of neurotactin described herein and the human form of neurotactin described herein is a chemokine-like domain which extends from about amino acid 22 to about amino acid 92.

- 4 -

Overall, the human form of neurotactin described herein is 67% identical at the amino acid level to the murine form of neurotactin described herein. The highest homology between the two forms is found in transmembrane and cytoplasmic domains, suggesting that both domains have important functional roles. High homology between the two forms is also found in the chemokine-like amino terminal region.

The invention features an isolated nucleic acid which encodes a neurotactin polypeptide. The nucleic acid can have the nucleotide sequence of, e.g., Fig. 1, SEQ ID NO:1 (murine), or Fig. 2, SEQ ID NO:3 (human). Preferably, the neurotactin polypeptide includes an amino acid sequence substantially identical to the amino acid sequence shown in Fig. 1 (SEQ ID NO:2) or the amino acid sequence shown in Fig. 2 (SEQ ID NO:4). Also considered within the scope of the invention are isolated nucleic acids that hybridize under stringent conditions to nucleic acids having the nucleotide sequence of, e.g., Fig. 1, SEQ ID NO:1 (murine), or Fig. 2, SEQ ID NO:3 (human). Substantially pure polypeptides encoded by nucleic acids that hybridize under stringent conditions to, e.g., SEQ ID NO:1 or SEQ ID NO:3, are also featured in the invention.

Preferred neurotactin polypeptides have a sequence which is substantially identical to that of a naturally occurring neurotactin polypeptide, e.g., the mature form of human neurotactin described herein.

By "isolated nucleic acid" is meant DNA that is not immediately contiguous with both of the coding sequences with which it is immediately contiguous (one on the 5' end and one on the 3' end) in the naturally occurring genome of the organism from which it is derived. Thus, a recombinant nucleic acid could include some or all of the 5' non-coding (e.g., promoter)

- 5 -

sequences which are immediately contiguous to the coding sequence. The term therefore includes, for example, a recombinant DNA which is incorporated into a vector; into an autonomously replicating plasmid or virus, such as a
5 retrovirus; or into the genomic DNA of a prokaryote or eukaryote, or which exists as a separate molecule (e.g., a cDNA or a genomic DNA fragment produced by PCR or restriction endonuclease treatment) independent of other sequences. It also includes a recombinant DNA which is
10 part of a hybrid gene encoding additional polypeptide sequence.

By "mature human neurotactin" is meant a polypeptide having the sequence shown in Fig. 2 (SEQ ID NO:4) from about amino acid 22 to amino acid 397.

15 Polypeptides substantially identical to mature human neurotactin have an amino acid sequence which is at least 85%, preferably 90%, and most preferably 95% or even 99% identical to the amino acid sequence of the neurotactin polypeptide of Fig. 2 (SEQ ID NO:4).

20 By "substantially identical" is meant a polypeptide or nucleic acid having a sequence that is at least 85%, preferably 90%, and more preferably 95% or more identical to the sequence of the reference amino acid or nucleic acid sequence. For polypeptides, the
25 length of the reference polypeptide sequence will generally be at least 16 amino acids, preferably at least 20 amino acids, more preferably at least 25 amino acids, and most preferably 35 amino acids. For nucleic acids, the length of the reference nucleic acid sequence will
30 generally be at least 50 nucleotides, preferably at least 60 nucleotides, more preferably at least 75 nucleotides, and most preferably 110 nucleotides.

Sequence identity can be measured using sequence analysis software (e.g., Sequence Analysis Software
35 Package of the Genetics Computer Group, University of

- 6 -

Wisconsin Biotechnology Center, 1710 University Avenue, Madison, WI 53705), with the default parameters therein.

In the case of polypeptide sequences which are less than 100% identical to a reference sequence, the 5 non-identical positions are preferably, but not necessarily, conservative substitutions for the reference sequence. Conservative substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine, and leucine; aspartic acid 10 and glutamic acid; asparagine and glutamine; serine and threonine; lysine and arginine; and phenylalanine and tyrosine.

Where a particular polypeptide is said to have a specific percent identity to a reference polypeptide of a 15 defined length, the percent identity is relative to the reference peptide. Thus, a peptide that is 50% identical to a reference polypeptide that is 100 amino acids long can be a 50 amino acid polypeptide that is completely identical to a 50 amino acid long portion of the 20 reference polypeptide. It might also be a 100 amino acid long polypeptide which is 50% identical to the reference polypeptide over its entire length. Of course, many other polypeptides will meet the same criteria.

Polypeptides corresponding to one or more domains 25 of neurotactin, e.g., the extracellular domain or the chemokine-like domain (about amino acid 22 to about amino acid 92 of a form of neurotactin described herein), are also within the scope of the invention. Preferred polypeptides are those which are soluble under normal 30 physiological conditions. Also within the invention are soluble fusion proteins in which a full-length form of neurotactin or a portion (e.g., one or more domains) thereof is fused to an unrelated protein or polypeptide (i.e., a fusion partner) to create a fusion protein.

- 7 -

The invention also features isolated nucleic acid sequences that encode a portion of neurotactin, including but not limited to the extracellular domain, the transmembrane domain, the cytoplasmic domain, the chemokine-like domain, and various functional domains of neurotactin. Also within the invention are nucleic acids encoding polypeptides corresponding to one or more domains of neurotactin, e.g., the extracellular domain or the chemokine-like domain. Preferred nucleic acids encode polypeptides that are soluble under normal physiological conditions. Also within the invention are nucleic acids encoding fusion proteins in which a portion of neurotactin or a portion (e.g., one or more domains) thereof is fused to an unrelated protein or polypeptide (i.e., a fusion partner) to create a fusion protein.

Encompassed within the invention are nucleic acid sequences that encode forms of neurotactin in which sequences are altered or deleted.

The nucleic acids of the invention include nucleic acids encoding mature neurotactin as well as neurotactin polypeptides fused to a polypeptide which facilitates secretion, e.g., a secretory sequence. Such a fused protein is typically referred to as a preprotein. The secretory sequence can be removed by the host cell to form the mature protein. Also within the invention are nucleic acids that encode mature neurotactin fused to a polypeptide sequence to produce an inactive proprotein. Proproteins can be converted into the active form of the protein by removal of the inactivating sequence.

The invention also encompasses nucleic acids that hybridize under stringent conditions to a nucleic acid encoding a neurotactin polypeptide. "Stringent conditions" means hybridization at 50°C in Church buffer (7% SDS, 0.5% NaHPO₄, 1 mM EDTA, 1% BSA) and washing at 50°C in 2X SSC. The hybridizing portion of the

- 8 -

hybridizing nucleic acids are preferably 20, 30, 50, or 70 bases long. Preferably, the hybridizing portion of the hybridizing nucleic acid is 95% or even 98% identical to the sequence of a portion of a nucleic acid encoding a neurotactin polypeptide. Hybridizing nucleic acids of the type described above can be used as a cloning probe, a primer (e.g., a PCR primer), or a diagnostic probe. Preferred hybridizing nucleic acids encode a polypeptide having some or all of the biological activities possessed by naturally-occurring neurotactin. Hybridizing nucleic acids can be splice variants encoded by one of the neurotactin genes described herein. Thus, they may encode a protein which is shorter or longer than the various forms of neurotactin described herein.

Hybridizing nucleic acids may also encode proteins which are related to neurotactin (e.g, proteins encoded by genes which include a portion having a relatively high degree of identity to a neurotactin gene described herein).

The invention also features substantially pure neurotactin polypeptides. Among the polypeptides encompassed within the invention are those corresponding to the extracellular domain, the transmembrane domain, the cytoplasmic domain, and various functional domains of neurotactin including the chemokine-like domain which corresponds to a domain extending from about amino acid 22 to about amino acid 92 of the form of murine neurotactin described herein and to a domain extending from about amino acid 22 to about amino acid 92 of the form of human neurotactin described herein.

The invention also encompasses polypeptides and nucleic acids whose sequences are substantially identical to that of a form of neurotactin described herein.

By "protein" and "polypeptide" is meant any chain of amino acids, regardless of length or post-

- 9 -

translational modification (e.g., glycosylation or phosphorylation).

By "substantially pure" is meant a preparation which is at least 60% by weight (dry weight) the compound of interest, i.e., a neurotactin polypeptide. Preferably the preparation is at least 75%, more preferably at least 90%, and most preferably at least 99%, by weight the compound of interest. Purity can be measured by any appropriate standard method, e.g., column chromatography, polyacrylamide gel electrophoresis, or HPLC analysis.

The term "nucleic acid" encompasses both RNA and DNA, including cDNA, genomic DNA, and synthetic (e.g., chemically synthesized) DNA. The nucleic acid may be double-stranded or single-stranded. Where single-stranded, the nucleic acid may be the sense strand or the antisense strand.

The polypeptides of the invention include, but are not limited to: recombinant polypeptides, natural polypeptides, and synthetic polypeptides as well as polypeptides which are preproteins or proproteins.

The polypeptides of the invention can be expressed fused to another polypeptide, e.g., a marker polypeptide or fusion partner. For example, the polypeptide can be fused to a hexa-histidine tag to facilitate purification of bacterially expressed protein or a hemagglutinin tag to facilitate purification of protein expressed in eukaryotic cells.

The invention features transformed cells harboring a nucleic acid encompassed by the invention. The invention also features vectors which include a nucleic acid of the invention which is properly positioned for expression. For example, the vector can be an expression vector, and can include one or more regulatory elements. Regulatory elements that can influence the expression of the nucleic acid inserted into the vector, such as

- 10 -

regulatory elements that direct tissue-specific expression, are well known to those of skill in the art. Examples of regulatory elements include the cytomegalovirus hCMV immediate early gene, the early
5 promoter of SV40 adenovirus, the late promoter of SV40 adenovirus, the lac system, the trp system, the TAC system, the TRC system, the major operator and promoter regions of phage lambda, the control regions of fd coat protein, the promoter for 3-phosphoglycerate kinase, the
10 promoters of acid phosphatase, and the promoters of the yeast α -mating factors. The vector can be a plasmid, or a virus, such as a retrovirus.

By "transformed cell" is meant a cell into which (or into an ancestor of which) has been introduced, by
15 means of recombinant DNA techniques, a DNA molecule encoding (as used herein) neurotactin polypeptide.

By "positioned for expression" is meant that the selected DNA molecule is positioned adjacent to one or more sequence elements which direct transcription and/or
20 translation of the sequence such that the sequence elements can control transcription and/or translation of the selected DNA (i.e., the selected DNA is operably associated with the sequence elements). Such operably associated elements can be used to facilitate the
25 production of a neurotactin polypeptide.

The invention also features purified antibodies which specifically bind a neurotactin protein or polypeptide.

By "purified antibody" is meant an antibody which
30 is at least 60%, by dry weight, free from the proteins and naturally-occurring organic molecules with which it is naturally associated. Preferably, the preparation is at least 75%, more preferably at least 90%, and most preferably at least 99%, by dry weight, antibody.

- 11 -

By "specifically binds" is meant an antibody that recognizes and binds to a particular antigen, e.g., neurotactin polypeptide, but which does not substantially recognize and bind to other molecules in a sample, e.g.,
5 a biological sample, which naturally includes neurotactin.

The invention also features antagonists and agonists of neurotactin. Antagonists can inhibit one or more of the functions of neurotactin. Suitable
10 antagonists can include large or small molecules, antibodies to neurotactin, and neurotactin polypeptides which compete with a native form of neurotactin. Agonists of neurotactin will enhance or facilitate one or more of the functions of neurotactin. Suitable agonists
15 can include, for example, large or small molecules and antibodies to neurotactin.

Also within the invention are nucleic acid molecules which can be used to interfere with neurotactin expression, e.g., antisense molecules and ribozymes.

20 The invention features substantially pure neurotactin polypeptides. In various preferred embodiments the polypeptide is soluble, the polypeptide includes the chemokine-like domain of neurotactin, the polypeptide includes the extracellular domain of
25 neurotactin, the polypeptide is at least 80% identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4, the polypeptide is at least 90% identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4, the polypeptide has an
30 amino acid sequence identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4, the polypeptide is at least 80% identical to the amino acid sequence from amino acid 22 to amino acid 397 in SEQ ID NO:4, the polypeptide is at least 90% identical to the
35 amino acid sequence from amino acid 22 to amino acid 397

- 12 -

in SEQ ID NO:4, and the polypeptide has an amino acid sequence identical to the amino acid sequence from amino acid 22 to amino acid 397 in SEQ ID NO:4.

The invention also features a substantially pure
5 polypeptide which includes a first portion and a second portion; the first portion includes a neurotactin polypeptide and the second portion includes an immunoglobulin constant region.

The invention also features a substantially pure
10 polypeptide which includes a first portion and a second portion; the first portion includes a neurotactin polypeptide the second portion includes a detectable marker. Examples of detectable markers include β -lactamase, chloramphenicol acetyltransferase (CAT),
15 adenosine deaminase (ADA), aminoglycoside phosphotransferase (neo^r, G418^r), dihydrofolate reductase (DHFR), hygromycin-B-phosphotransferase (HPH), thymidine kinase (TK), β -galactosidase, and xanthine guanine phosphoribosyl-transferase (XGPRT).

20 In another aspect the invention features a recombinant nucleic acid encoding a neurotactin polypeptide. In various preferred embodiments the nucleic acid encodes a soluble neurotactin polypeptide, the nucleic acid encodes the chemokine-like domain of
25 neurotactin, and the nucleic acid encodes the extracellular domain of neurotactin.

The invention also features a nucleic acid encoding a hybrid polypeptide. This hybrid polypeptide includes a first portion and a second portion; the first
30 portion includes a neurotactin polypeptide; the second portion comprising an immunoglobulin constant region.

The invention also features a cell which harbors a recombinant nucleic acid encoding a neurotactin polypeptide; a vector which includes a nucleic acid
35 encoding a neurotactin polypeptide.

- 13 -

In another aspect, the invention features an antibody which selectively binds to a neurotactin polypeptide. In a preferred embodiment, the antibody is a monoclonal antibody.

5 The invention also features a pharmaceutical composition which includes a neurotactin polypeptide.

 The invention features a method for detecting inflammation. This method includes: (a) obtaining a biological sample; (b) contacting the sample with an
10 antibody which selectively binds a neurotactin polypeptide; and (c) determining the amount of the antibody selectively bound to said biological sample as a measure of inflammation.

 In another aspect, the invention features a method
15 for treating inflammation in a patient which includes administering to the patient an inhibitor of neurotactin. Preferably, the inhibitor is an antibody which selectively binds to neurotactin.

 The invention also features a method for
20 inhibiting proliferation of progenitor cells in a patient. The method includes administering to the patient a substantially pure neurotactin polypeptide capable of inhibiting progenitor cell proliferation. The invention also includes a method of suppressing
25 proliferation of an actively dividing myeloid cell. This method includes contacting the cell with an effective amount of a neurotactin polypeptide that is capable of inhibiting proliferation of myeloid cells.

 The invention also features an adjunctive method
30 for use with chemotherapy or radiation therapy in a patient. The method includes: administering an effective amount of a neurotactin polypeptide to the patient, and administering chemotherapy or radiation therapy to the patient in conjunction with administration
35 of the neurotactin polypeptide. By "adjunctive method"

- 14 -

means administration before, during, or after chemotherapy or radiation therapy.

The invention also features a method of treating a hyperproliferative myeloid disease in a patient. The method includes administering to the patient an effective amount of a neurotactin polypeptide. In preferred embodiments, the disease is chronic myelogenous leukemia, polycythemia vera, and a hypermegakaryocytopoietic disorder.

10 The invention features a substantially pure protein which functionally interacts with neurotactin and a nucleic acid encoding a protein which functionally interacts with neurotactin.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are described herein. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be limiting

25 Other features and advantages of the invention will be apparent from the following detailed descriptions, and from the claims.

Brief Description of the Drawings

Fig. 1 is a depiction of the nucleotide (SEQ ID NO:1) and amino acid (SEQ ID NO:2) sequences of a form of murine neurotactin, including a putative signal sequence. The unique arrangement between the C32 and C36 is underlined. Also underlined are R337 and R338, a

- 15 -

potential enzyme cleavage site, and a potential membrane spanning domain from A340 to Y360.

Fig. 2 is a representation of the nucleotide (SEQ ID NO:3) and amino acid (SEQ ID NO:4) sequence of a form of human neurotactin, including a putative signal sequence. The unique arrangement between the C32 and C36 is underlined. Also underlined are the conserved R339 and R340, a potential enzyme cleavage site, and a potential membrane spanning domain from A342 to Y362.

Fig. 3 is a representation of the sequence alignment of full-length human and mouse neurotactin. In this representation, a vertical line between the two aligned sequences indicates an exact match, a single dot between the two aligned sequences indicates a conservative substitution, a pair of dots between the two aligned sequences indicates a very conservative substitution, and a series of dots within a sequence indicates a gap introduced to maximize alignment. Fig. 4 is a comparison of the primary structure of neurotactin to that of known chemokine subfamilies.

Fig. 5 is a graph illustrating the results of a neutrophil chemotaxis assay.

Detailed Description

Neurotactin, described for the first time herein, is a novel chemokine that plays a role in inflammation, particularly inflammation of the brain.

Neurotactin mediates chemotaxis of specific cell types and is likely to induce release of inflammatory mediators. As a consequence, neurotactin may enhance leukocyte infiltration through the endothelial cell wall and have effect on microglial cells. Neurotactin, like certain other proteins, may have a functional secreted form as well as a functional membrane-bound form.

- 16 -

While it is clear that neurotactin is a chemokine, it is also clear that neurotactin is an example of a novel class of chemokines.

First, neurotactin has three spacer amino acids
5 between the first two cysteines (CXXXC), compared with none or one in the CC or CXC chemokines, respectively.

Second, neurotactin has an unusual expression pattern. As with other chemokines, neurotactin can be upregulated in bone marrow stromal cells, endothelial
10 cells, and fibroblasts when treated with LPS and PMA. However, unlike other chemokines, neurotactin is highly expressed in normal brain, which suggests involvement in brain function.

Third, the length of the predicted neurotactin
15 protein, 395 amino acids for the murine form described herein and 397 amino acids for the human form described herein, is considerably longer than the majority of known chemokines. Further, the portions of murine and human neurotactin after the first 92 amino acids of the mature
20 protein do not bear significant resemblance to any presently known, sequenced protein. The first 92 acids of both the murine and human forms of neurotactin described herein have greater than 40% identity to murine MCP-1 and human MCP-1, respectively.

25 Fourth, the neurotactin gene maps to a different chromosomal location than the three known classes of mouse chemokines. It is located on human chromosome 16 and mouse chromosome 8. The murine CC, CXC, and lymphotactin chemokines have been mapped to chromosomes
30 11, 5, and 1 respectively.

Taken together, these facts demonstrate that neurotactin represents a new class of chemokine, referred to herein as the δ class.

Fig. 4 shows the primary structure of the murine
35 and human forms of neurotactin, as well as the primary

- 17 -

structures of human IL-8, human MCP-1, and human lymphotactin. Also shown in this figure are comparisons between the proteins. As can be seen from this figure, the primary structure of the first 92 amino acids of all
5 five proteins are similar, whereas the two forms of neurotactin include an extracellular linker domain, a transmembrane domain, and a cytoplasmic domain not found in the other chemokines. The extracellular linker domain is the domain between the chemotactin-like domain and the
10 transmembrane domain. The extracellular domain extends from about amino acid 93 to about amino acid 341 of the human form of neurotactin described herein and from about amino acid 93 to about amino acid 339 of the murine form of neurotactin described herein.

15 The two neighboring arginines adjacent the transmembrane region (amino acids 339 and 340 in the human form; amino acids 337 and 338 in the murine form) provide for the possibility of processing these proteins with proteolytic enzymes to detach them from the cell
20 membrane.

Neurotactin Proteins and Polypeptides

Neurotactin proteins and polypeptides and neurotactin fusion proteins can be prepared for a wide range of uses including, but not limited to, generation
25 of antibodies, preparation of reagents for diagnostic assays, identification of other molecules involved in inflammation (particularly brain inflammation), preparation of reagents for use in screening assays for inflammatory modulators, and preparation of therapeutic
30 agents for treatment of inflammation-related disorders.

Fig. 1 shows the amino acid sequence of a form of murine neurotactin (SEQ ID NO:2). The domain from amino acid 1 to approximately amino acid 21 (in italics) forms a putative signal sequence. This putative signal

- 18 -

sequence is followed by an apparent extracellular domain of approximately 318 amino acids extending from about amino acid 22 to about amino acid 339; a transmembrane domain of approximately 21 amino acids extending from
5 about amino acid 340 to about amino acid 360; and a cytoplasmic domain of approximately 35 amino acids extending from about amino acid 361 to amino acid 395.

Fig. 2 shows the amino acid sequence of a form of human neurotactin (SEQ ID NO: 4). The domain from amino
10 acid 1 to approximately amino acid 21 (in italics) forms a putative signal sequence. This putative signal sequence is followed by an apparent extracellular domain of approximately 321 amino acids extending from about amino acid 22 to about amino acid 341; a transmembrane
15 domain of approximately 22 amino acids extending from about amino acid 342 to about amino acid 362; and a cytoplasmic domain of approximately 35 amino acids extending from about amino acid 363 to amino acid 397.

The invention encompasses, but is not limited to,
20 neurotactin proteins and polypeptides that are functionally related to neurotactin encoded by the nucleotide sequence of Fig. 1 (murine, SEQ ID NO:1) or Fig. 2 (human, SEQ ID NO:3). Functionally related proteins and polypeptides include any protein or
25 polypeptide sharing a functional characteristic with neurotactin, e.g., the ability to affect proliferation, differentiation, survival, apoptosis, or activation of a cell type whose proliferation, differentiation, survival, apoptosis, or activation is affected by neurotactin.
30 Such functionally related neurotactin polypeptides include, but are not limited to, additions or substitutions of amino acid residues within the amino acid sequence encoded by the neurotactin sequences described herein which result in a silent change, thus
35 producing a functionally equivalent gene product. Amino

- 19 -

acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

- 5 For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged
10 (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

- While random mutations can be made to neurotactin DNA (using random mutagenesis techniques well known to
15 those skilled in the art) and the resulting mutant neurotactin proteins can be tested for activity, site-directed mutations of the neurotactin coding sequence can be engineered (using site-directed mutagenesis techniques well known to those skilled in the art) to generate
20 mutant neurotactins with increased function, e.g., greater stimulation of cell proliferation, or decreased function, e.g., lesser stimulation of cell proliferation.

- To design functionally related and functionally variant neurotactin polypeptides, it is useful to
25 distinguish between conserved positions and variable positions. Fig. 4 shows an alignment between the amino acid sequence of human neurotactin and murine neurotactin, which can be used to determine the conserved and variable amino acid positions.

- 30 To preserve neurotactin function, it is preferable that conserved residues are not altered. Moreover, alteration of non-conserved residues are preferably conservative alterations, e.g., a basic amino acid is replaced by a different basic amino acid. To produce
35 altered function variants, it is preferable to make non-

- 20 -

conservative changes at variable and/or conserved positions. Deletions at conserved and variable positions can also be used to create altered function variants.

Other mutations to the neurotactin coding sequence
5 can be made to generate neurotactins that are better suited for expression, scale up, etc. in a selected host cell. For example, N-linked glycosylation sites can be altered or eliminated to achieve, for example, expression of a homogeneous product that is more easily recovered
10 and purified from yeast hosts which are known to hyperglycosylate N-linked sites. To this end, a variety of amino acid substitutions at one or both of the first or third amino acid positions of any one or more of the glycosylation recognition sequences which occur (in N-X-S
15 or N-X-T), and/or an amino acid deletion at the second position of any one or more of such recognition sequences, will prevent glycosylation at the modified tripeptide sequence. (See , e.g., Miyajima et al., *EMBO J.* 5:1193, 1986).

20 Preferred neurotactin polypeptides are those neurotactin polypeptides, or variants thereof, which stimulate chemotaxis of neutrophils. In determining whether a particular neurotactin polypeptide or variant thereof stimulates chemotaxis of neutrophils, one can use
25 any standard neutrophil chemotaxis assay. One preferred assay is the chemotaxis assay described herein. Preferred neurotactin polypeptides and variants have 20%, 40%, 50%, 75%, 80%, or even 90% of the activity of the full-length, mature human form of neurotactin described
30 herein. Such comparisons are generally based on equal concentrations of the molecules being compared. The comparison can also be based on the amount of protein or polypeptide required to reach 50% of the maximal stimulation obtainable.

- 21 -

Polypeptides corresponding to one or more domains of neurotactin, e.g., the extracellular domain and the chemokine-like domain, are also within the scope of the invention. Preferred polypeptides are those which are
5 soluble under normal physiological conditions. Also within the invention are fusion proteins in which a portion (e.g., one or more domains) of neurotactin is fused to an unrelated protein or polypeptide (i.e., a fusion partner) to create a fusion protein. The fusion
10 partner can be a moiety selected to facilitate purification, detection, or solubilization, or to provide some other function. Fusion proteins are generally produced by expressing a hybrid gene in which a nucleotide sequence encoding all or a portion of
15 neurotactin is joined in-frame to a nucleotide sequence encoding the fusion partner. Fusion partners include, but are not limited to, the constant region of an immunoglobulin (IgFc). A fusion protein in which a neurotactin polypeptide is fused to IgFc can be more
20 stable and have a longer half-life in the body than the neurotactin polypeptide on its own.

Also within the scope of the invention are various soluble forms of neurotactin. For example, the entire extracellular domain of neurotactin or a portion thereof
25 can be expressed on its own or fused to a solubilization partner, e.g., an immunoglobulin.

The invention also features neurotactin polypeptides that can inhibit proliferation of progenitor cells. Such polypeptides are can be used to protect
30 progenitor cells from the effects of chemotherapy and/or radiation therapy. Any convenient *in vitro* or *in vivo* assay can be used to determine whether a selected neurotactin polypeptide or variant thereof inhibits progenitor cell proliferation and is thus likely to be a
35 suitable chemoprotective agent. Suitable *in vitro*

- 22 -

assays include those described by Gentile et al. (U.S. Patents Nos. 5,149,544 and 5,294,544). In addition, inhibition of progenitor cell proliferation can be tested using an *in vivo* assay. A suitable murine model for
5 assessing progenitor cell proliferation has been described by Cooper et al. (*Exp. Hematol.* 22:186, 1994). The results of this *in vivo* model, together with the *in vitro* assay results, are predictive of the efficacy of the tested molecules in treating patients, e.g., humans.

10 In general, neurotactin proteins according to the invention can be produced by transformation (transfection, transduction, or infection) of a host cell with all or part of a neurotactin-encoding DNA fragment (e.g., the cDNA described herein) in a suitable
15 expression vehicle. Suitable expression vehicles include: plasmids, viral particles, and phage. For insect cells, baculovirus expression vectors are suitable. The entire expression vehicle, or a part thereof, can be integrated into the host cell genome. In
20 some circumstances, it is desirable to employ an inducible expression vector, e.g., the LACSWITCH™ Inducible Expression System (Stratagene, LaJolla, CA).

Those skilled in the field of molecular biology will understand that any of a wide variety of expression
25 systems can be used to provide the recombinant protein. The precise host cell used is not critical to the invention. The neurotactin protein can be produced in a prokaryotic host (e.g., *E. coli* or *B. subtilis*) or in a eukaryotic host (e.g., *Saccharomyces* or *Pichia*; mammalian
30 cells, e.g., COS, NIH 3T3, CHO, BHK, 293, or HeLa cells; or insect cells).

Proteins and polypeptides can also be produced by plant cells. For plant cells viral expression vectors (e.g., cauliflower mosaic virus and tobacco mosaic virus)
35 and plasmid expression vectors (e.g., Ti plasmid) are

- 23 -

suitable. Such cells are available from a wide range of sources (e.g., the American Type Culture Collection, Rockland, MD; also, see, e.g., Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York, 1994). The methods of transformation or transfection and the choice of expression vehicle will depend on the host system selected. Transformation and transfection methods are described, e.g., in Ausubel et al. (*Current Protocols in Molecular Biology*, John Wiley & Sons, New York, 1994); expression vehicles may be chosen from those provided, e.g., in *Cloning Vectors: A Laboratory Manual* (P.H. Pouwels et al., 1985, Supp. 1987).

The host cells harboring the expression vehicle can be cultured in conventional nutrient media adapted as need for activation of a chosen gene, repression of a chosen gene, selection of transformants, or amplification of a chosen gene.

One preferred expression system is the mouse 3T3 fibroblast host cell transfected with a pMAMneo expression vector (Clontech, Palo Alto, CA). pMAMneo provides an RSV-LTR enhancer linked to a dexamethasone-inducible MMTV-LTR promotor, an SV40 origin of replication which allows replication in mammalian systems, a selectable neomycin gene, and SV40 splicing and polyadenylation sites. DNA encoding a neurotactin protein would be inserted into the pMAMneo vector in an orientation designed to allow expression. The recombinant neurotactin protein would be isolated as described below. Other preferable host cells that can be used in conjunction with the pMAMneo expression vehicle include COS cells and CHO cells (ATCC Accession Nos. CRL 1650 and CCL 61, respectively).

Neurotactin polypeptides can be produced as fusion proteins. For example, the expression vector pUR278

- 24 -

(Ruther et al., *EMBO J.* 2:1791, 1983), can be used to create lacZ fusion proteins. The pGEX vectors can be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can be easily purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned target gene product can be released from the GST moiety.

In an insect cell expression system, *Autographa californica* nuclear polyhydrosis virus (AcNPV), which grows in *Spodoptera frugiperda* cells, is used as a vector to express foreign genes. A neurotactin coding sequence can be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter, e.g., the polyhedrin promoter. Successful insertion of a gene encoding a neurotactin polypeptide or protein will result in inactivation of the polyhedrin gene and production of non-occluded recombinant virus (i.e., virus lacking the proteinaceous coat encoded by the polyhedrin gene). These recombinant viruses are then used to infect *spodoptera frugiperda* cells in which the inserted gene is expressed (see, e.g., Smith et al., *J. Virol.* 46:584, 1983; Smith, U.S. Patent No. 4,215,051).

In mammalian host cells, a number of viral-based expression systems can be utilized. In cases where an adenovirus is used as an expression vector, the neurotactin nucleic acid sequence can be ligated to an adenovirus transcription/translation control complex, e.g., the late promoter and tripartite leader sequence. This chimeric gene can then be inserted into the adenovirus genome by *in vitro* or *in vivo* recombination.

- 25 -

Insertion into a non-essential region of the viral genome (e.g., region E1 or E3) will result in a recombinant virus that is viable and capable of expressing a neurotactin gene product in infected hosts (see, e.g.,
5 Logan, *Proc. Natl. Acad. Sci. USA* 81:3655, 1984).

Specific initiation signals may also be required for efficient translation of inserted nucleic acid sequences. These signals include the ATG initiation codon and adjacent sequences. In cases where an entire
10 native neurotactin gene or cDNA, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. In other cases, exogenous translational control signals,
15 including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and
20 initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators (Bittner et al., *Methods in Enzymol.* 153:516, 1987).

25 In addition, a host cell may be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in a specific, desired fashion. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein
30 products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems can be chosen to
35 ensure the correct modification and processing of the

- 26 -

foreign protein expressed. To this end, eukaryotic host cells that possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product can be used. Such
5 mammalian host cells include, but are not limited to, CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, and in particular, choroid plexus cell lines.

Alternatively, a neurotactin protein can be produced by a stably-transfected mammalian cell line. A
10 number of vectors suitable for stable transfection of mammalian cells are available to the public, see, e.g., Pouwels et al. (*supra*); methods for constructing such cell lines are also publicly available, e.g., in Ausubel et al. (*supra*). In one example, cDNA encoding the
15 neurotactin protein is cloned into an expression vector that includes the dihydrofolate reductase (DHFR) gene. Integration of the plasmid and, therefore, the neurotactin protein-encoding gene into the host cell chromosome is selected for by including 0.01-300 μ M
20 methotrexate in the cell culture medium (as described in Ausubel et al., *supra*). This dominant selection can be accomplished in most cell types.

Recombinant protein expression can be increased by DHFR-mediated amplification of the transfected gene.
25 Methods for selecting cell lines bearing gene amplifications are described in Ausubel et al. (*supra*); such methods generally involve extended culture in medium containing gradually increasing levels of methotrexate. DHFR-containing expression vectors commonly used for this
30 purpose include pCVSEII-DHFR and pAdD26SV(A) (described in Ausubel et al., *supra*). Any of the host cells described above or, preferably, a DHFR-deficient CHO cell line (e.g., CHO DHFR⁻ cells, ATCC Accession No. CRL 9096) are among the host cells preferred for DHFR selection of

- 27 -

a stably-transfected cell line or DHFR-mediated gene amplification.

A number of other selection systems can be used, including but not limited to the herpes simplex virus thymidine kinase, hypoxanthine-guanine phosphoribosyl-transferase, and adenine phosphoribosyltransferase genes can be employed in tk, hgp^{rt}, or ap^{rt} cells, respectively. In addition, gpt, which confers resistance to mycophenolic acid (Mulligan et al., *Proc. Natl. Acad. Sci. USA* 78:2072, 1981); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin et al., *J. Mol. Biol.* 150:1, 1981); and hyg^{ro}, which confers resistance to hygromycin (Santerre et al., *Gene* 30:147, 1981), can be used.

Alternatively, any fusion protein can be readily purified by utilizing an antibody specific for the fusion protein being expressed. For example, a system described in Janknecht et al., *Proc. Natl. Acad. Sci. USA*, 88:8972 (1981), allows for the ready purification of non-denatured fusion proteins expressed in human cell lines. In this system, the gene of interest is subcloned into a vaccinia recombination plasmid such that the gene's open reading frame is translationally fused to an amino-terminal tag consisting of six histidine residues. Extracts from cells infected with recombinant vaccinia virus are loaded onto Ni²⁺ nitriloacetic acid-agarose columns, and histidine-tagged proteins are selectively eluted with imidazole-containing buffers.

Alternatively, neurotactin or a portion thereof, can be fused to an immunoglobulin Fc domain. Such a fusion protein can be readily purified using a protein A column. Moreover, such fusion proteins permit the production of a dimeric form of a neurotactin polypeptide having increased stability *in vivo*.

- 28 -

Neurotactin proteins and polypeptides can also be expressed in transgenic animals. Animals of any species, including, but not limited to, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, goats, and non-human
5 primates, e.g., baboons, monkeys, and chimpanzees, can be used to generate neurotactin-expressing transgenic animals.

Any technique known in the art can be used to introduce a neurotactin transgene into animals to produce
10 the founder lines of transgenic animals. Such techniques include, but are not limited to, pronuclear microinjection (U.S. Pat. No. 4,873,191); retrovirus mediated gene transfer into germ lines (Van der Putten et al., *Proc. Natl. Acad. Sci., USA* 82:6148, 1985); gene
15 targeting into embryonic stem cells (Thompson et al., *Cell* 56:313, 1989); and electroporation of embryos (Lo, *Mol. Cell. Biol.* 3:1803, 1983).

The present invention provides for transgenic animals that carry the neurotactin transgene in all their
20 cells, as well as animals that carry the transgene in some, but not all of their cells, i.e., mosaic animals. The transgene can be integrated as a single transgene or in concatamers, e.g., head-to-head tandems or head-to-tail tandems. The transgene can also be selectively
25 introduced into and activated in a particular cell type (Lasko et al., *Proc. Natl. Acad. Sci. USA* 89:6232, 1992). The regulatory sequences required for such a cell-type specific activation will depend upon the particular cell type of interest, and will be apparent to those of skill
30 in the art.

When it is desired that the neurotactin transgene be integrated into the chromosomal site of the endogenous neurotactin gene, gene targeting is preferred. Briefly, when such a technique is to be used, vectors containing
35 some nucleotide sequences homologous to an endogenous

- 29 -

neurotactin gene are designed for the purpose of integrating, via homologous recombination with chromosomal sequences, into and disrupting the function of the nucleotide sequence of the endogenous gene. The transgene also can be selectively introduced into a particular cell type, thus inactivating the endogenous neurotactin gene in only that cell type (Gu et al., *Science* 265:103, 1984). The regulatory sequences required for such a cell-type specific inactivation will depend upon the particular cell type of interest, and will be apparent to those of skill in the art.

Once transgenic animals have been generated, the expression of the recombinant neurotactin gene can be assayed utilizing standard techniques. Initial screening may be accomplished by Southern blot analysis or PCR techniques to analyze animal tissues to assay whether integration of the transgene has taken place. The level of mRNA expression of the transgene in the tissues of the transgenic animals may also be assessed using techniques which include, but are not limited to, Northern blot analysis of tissue samples obtained from the animal, *in situ* hybridization analysis, and RT-PCR. Samples of neurotactin gene-expressing tissue, also can be evaluated immunocytochemically using antibodies specific for the neurotactin transgene product.

Once the recombinant neurotactin protein is expressed, it is isolated. Secreted forms can be isolated from the culture media, while non-secreted forms must be isolated from the host cells. Proteins can be isolated by affinity chromatography. In one example, an anti-neurotactin protein antibody (e.g., produced as described herein) is attached to a column and used to isolate the neurotactin protein. Lysis and fractionation of neurotactin protein-harboring cells prior to affinity chromatography can be performed by standard methods (see,

- 30 -

e.g., Ausubel et al., *supra*). Alternatively, a neurotactin fusion protein, for example, a neurotactin-maltose binding protein, a neurotactin- β -galactosidase, or a neurotactin-trpE fusion protein, can be constructed
5 and used for neurotactin protein isolation (see, e.g., Ausubel et al., *supra*; New England Biolabs, Beverly, MA).

Once isolated, the recombinant protein can, if desired, be further purified, e.g., by high performance liquid chromatography using standard techniques (see,
10 e.g., Fisher, *Laboratory Techniques In Biochemistry And Molecular Biology*, eds., Work and Burdon, Elsevier, 1980).

Polypeptides of the invention, particularly short neurotactin fragments, can also be produced by chemical
15 synthesis (e.g., by the methods described in *Solid Phase Peptide Synthesis*, 2nd ed., 1984 The Pierce Chemical Co., Rockford, IL).

These general techniques of polypeptide expression and purification can also be used to produce and isolate
20 useful neurotactin fragments or analogs (described herein).

The invention also features proteins which interact with neurotactin and are involved in the function of neurotactin. Also included in the invention
25 are the genes encoding these interacting proteins. Interacting proteins can be identified using methods known to those skilled in the art. One suitable method is the "two-hybrid system," detects protein interactions in vivo (Chien et al., *Proc. Natl. Acad. Sci. USA*
30 88:9578, 1991). A kit for practicing this method is available from Clontech (Palo Alto, CA).

Anti-Neurotactin Antibodies

Human neurotactin proteins and polypeptides (or immunogenic fragments or analogs) can be used to raise

- 31 -

antibodies useful in the invention; such polypeptides can be produced by recombinant or peptide synthetic techniques (see, e.g., *Solid Phase Peptide Synthesis*, *supra*; Ausubel et al., *supra*). In general, the peptides
5 can be coupled to a carrier protein, such as KLH, as described in Ausubel et al., *supra*, mixed with an adjuvant, and injected into a host mammal. Antibodies can be purified by peptide antigen affinity chromatography.

10 In particular, various host animals can be immunized by injection with a neurotactin protein or polypeptide. Host animals include rabbits, mice, guinea pigs, and rats. Various adjuvants can be used to increase the immunological response, depending on the
15 host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and
20 potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium parvum*. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

Antibodies within the invention include monoclonal
25 antibodies, polyclonal antibodies, humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, and molecules produced using a Fab expression library.

Monoclonal antibodies, which are homogeneous
30 populations of antibodies to a particular antigen, can be prepared using the neurotactin proteins described above and standard hybridoma technology (see, e.g., Kohler et al., *Nature* 256:495, 1975; Kohler et al., *Eur. J. Immunol.* 6:511, 1976; Kohler et al., *Eur. J. Immunol.*
35 6:292, 1976; Hammerling et al., *In Monoclonal Antibodies*

- 32 -

and *T Cell Hybridomas*, Elsevier, NY, 1981; Ausubel et al., *supra*).

In particular, monoclonal antibodies can be obtained by any technique that provides for the
5 production of antibody molecules by continuous cell lines in culture such as described in Kohler et al., *Nature* 256:495, 1975, and U.S. Patent No. 4,376,110; the human B-cell hybridoma technique (Kosbor et al., *Immunology Today* 4:72, 1983; Cole et al., *Proc. Natl. Acad. Sci. USA*
10 80:2026, 1983), and the EBV-hybridoma technique (Cole et al., *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96, 1983). Such antibodies can be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing
15 the mAb of this invention may be cultivated *in vitro* or *in vivo*. The ability to produce high titers of mAbs *in vivo* makes this the presently preferred method of production.

Once produced, polyclonal or monoclonal antibodies
20 are tested for specific neurotactin recognition by Western blot or immunoprecipitation analysis by standard methods, e.g., as described in Ausubel et al., *supra*. Antibodies that specifically recognize and bind to neurotactin are useful in the invention. For example,
25 such antibodies can be used in an immunoassay to monitor the level of neurotactin produced by a mammal (for example, to determine the amount or subcellular location of neurotactin).

Preferably, antibodies of the invention are
30 produced using fragments of the neurotactin protein which lie outside highly conserved regions and appear likely to be antigenic, by criteria such as high frequency of charged residues. In one specific example, such fragments are generated by standard techniques of PCR,
35 and are then cloned into the pGEX expression vector

- 33 -

(Ausubel et al., *supra*). Fusion proteins are expressed in *E. coli* and purified using a glutathione agarose affinity matrix as described in Ausubel, et al., *supra*.

In some cases it may be desirable to minimize the
5 potential problems of low affinity or specificity of antisera. In such circumstances, two or three fusions can be generated for each protein, and each fusion can be injected into at least two rabbits. Antisera can be raised by injections in a series, preferably including at
10 least three booster injections.

Antisera is also checked for its ability to immunoprecipitate recombinant neurotactin proteins or control proteins, such as glucocorticoid receptor, CAT, or luciferase.

15 The antibodies can be used, for example, in the detection of the neurotactin in a biological sample as part of a diagnostic assay. Antibodies also can be used in a screening assay to measure the effect of a candidate compound on expression or localization of neurotactin.
20 Additionally, such antibodies can be used in conjunction with the gene therapy techniques described to, for example, evaluate the normal and/or engineered neurotactin-expressing cells prior to their introduction into the patient. Such antibodies additionally can be
25 used in a method for inhibiting abnormal neurotactin activity.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., *Proc. Natl. Acad. Sci. USA* 81:6851, 1984; Neuberger et
30 al., *Nature* 312:604, 1984; Takeda et al., *Nature* 314:452, 1984) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is
35 a molecule in which different portions are derived from

- 34 -

different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region.

Alternatively, techniques described for the
5 production of single chain antibodies (U.S. Patent 4,946,778; and U.S. Patents 4,946,778 and 4,704,692) can be adapted to produce single chain antibodies against a neurotactin protein or polypeptide. Single chain
10 chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments that recognize and bind to specific epitopes can be generated by known techniques. For example, such fragments include but are not limited
15 to F(ab')₂ fragments that can be produced by pepsin digestion of the antibody molecule, and Fab fragments that can be generated by reducing the disulfide bridges of F(ab')₂ fragments. Alternatively, Fab expression libraries can be constructed (Huse et al., *Science*
20 246:1275, 1989) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Antibodies to the neurotactin can, in turn, be used to generate anti-idiotypic antibodies that resemble a portion of neurotactin using techniques well known to
25 those skilled in the art (see, e.g., Greenspan et al., *FASEB J.* 7:437, 1993; Nissinoff, *J. Immunol.* 147:2429, 1991). For example, antibodies that bind to neurotactin and competitively inhibit the binding of a ligand of neurotactin can be used to generate anti-idiotypes that
30 resemble a ligand binding domain of neurotactin and, therefore, bind and neutralize a ligand of neurotactin. Such neutralizing anti-idiotypic antibodies or Fab fragments of such anti-idiotypic antibodies can be used in therapeutic regimens.

- 35 -

Reducing Neurotactin Expression

- In alternate embodiments, anti-inflammation therapy can be designed to reduce the level of endogenous neurotactin gene expression, e.g., using antisense or
- 5 ribozyme approaches to inhibit or prevent translation of neurotactin mRNA transcripts; triple helix approaches to inhibit transcription of the neurotactin gene; or targeted homologous recombination to inactivate or "knock out" the neurotactin gene or its endogenous promoter.
- 10 Because the neurotactin gene is expressed in the brain, including the choroid plexus and arcuate nucleus, delivery techniques should be preferably designed to cross the blood-brain barrier (see, e.g., PCT WO89/10134). Alternatively, the antisense, ribozyme, or
- 15 DNA constructs described herein could be administered directly to the site containing the target cells; e.g., brain, heart, kidney, lung, uterus, endothelial cells, fibroblasts, and bone marrow stromal cells.

Antisense Nucleic Acids

- 20 Antisense approaches involve the design of oligonucleotides (either DNA or RNA) that are complementary to neurotactin mRNA. The antisense oligonucleotides bind to the complementary neurotactin mRNA transcripts and prevent translation. Absolute
- 25 complementarity, although preferred, is not required. A sequence "complementary" to a portion of an RNA, as referred to herein, means a sequence having sufficient complementarity to be able to hybridize with the RNA, forming a stable duplex; in the case of double-stranded
- 30 antisense nucleic acids, a single strand of the duplex DNA may be tested, or triplex formation may be assayed. The ability to hybridize will depend on both the degree of complementarity and the length of the antisense nucleic acid. Generally, the longer the hybridizing

- 36 -

nucleic acid, the more base mismatches with an RNA it may contain and still form a stable duplex (or triplex, as the case may be). One skilled in the art can ascertain a tolerable degree of mismatch by use of standard
5 procedures to determine the melting point of the hybridized complex.

Oligonucleotides that are complementary to the 5' end of the message, e.g., the 5' untranslated sequence up to and including the AUG initiation codon, should work
10 most efficiently at inhibiting translation. However, sequences complementary to the 3' untranslated sequences of mRNAs recently have been shown to be effective at inhibiting translation of mRNAs as well (Wagner, *Nature* 372:333, 1984). Thus, oligonucleotides complementary to
15 either the 5'- or 3'-non-translated, non-coding regions of the neurotactin gene, e.g., the human gene shown in Fig. 2, could be used in an antisense approach to inhibit translation of endogenous neurotactin mRNA.
Oligonucleotides complementary to the 5' untranslated
20 region of the mRNA should include the complement of the AUG start codon.

Antisense oligonucleotides complementary to mRNA coding regions are less efficient inhibitors of translation but could be used in accordance with the
25 invention. Whether designed to hybridize to the 5'-, 3'-, or coding region of neurotactin mRNA, antisense nucleic acids should be at least six nucleotides in length, and are preferably oligonucleotides ranging from 6 to about 50 nucleotides in length. In specific aspects the
30 oligonucleotide is at least 10 nucleotides, at least 17 nucleotides, at least 25 nucleotides or at least 50 nucleotides.

Regardless of the choice of target sequence, it is preferred that *in vitro* studies are first performed to
35 quantitate the ability of the antisense oligonucleotide

- 37 -

to inhibit gene expression. It is preferred that these studies utilize controls that distinguish between antisense gene inhibition and nonspecific biological effects of oligonucleotides. It is also preferred that
5 these studies compare levels of the target RNA or protein with that of an internal control RNA or protein. Additionally, it is envisioned that results obtained using the antisense oligonucleotide are compared with those obtained using a control oligonucleotide. It is
10 preferred that the control oligonucleotide is of approximately the same length as the test oligonucleotide and that the nucleotide sequence of the oligonucleotide differs from the antisense sequence no more than is necessary to prevent specific hybridization to the target
15 sequence.

The oligonucleotides can be DNA or RNA or chimeric mixtures or derivatives or modified versions thereof, single-stranded or double-stranded. The oligonucleotide can be modified at the base moiety, sugar moiety, or
20 phosphate backbone, for example, to improve stability of the molecule, hybridization, etc. The oligonucleotide may include other appended groups such as peptides (e.g., for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (as
25 described, e.g., in Letsinger et al., *Proc. Natl. Acad. Sci. USA* 86:6553, 1989; Lemaitre et al., *Proc. Natl. Acad. Sci. USA* 84:648, 1987; PCT Publication No. WO 88/09810) or the blood-brain barrier (see, e.g., PCT Publication No. WO 89/10134), or hybridization-triggered
30 cleavage agents (see, e.g., Krol et al., *BioTechniques* 6:958, 1988), or intercalating agents (see, e.g., Zon, *Pharm. Res.* 5:539, 1988). To this end, the oligonucleotide can be conjugated to another molecule, e.g., a peptide, hybridization triggered cross-linking

- 38 -

agent, transport agent, or hybridization-triggered cleavage agent.

The antisense oligonucleotide may comprise at least one modified base moiety which is selected from the group including, but not limited to, 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xantine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 2-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine.

The antisense oligonucleotide may also comprise at least one modified sugar moiety selected from the group including, but not limited to, arabinose, 2-fluoroarabinose, xylulose, and hexose.

In yet another embodiment, the antisense oligonucleotide comprises at least one modified phosphate backbone selected from the group consisting of a phosphorothioate, a phosphorodithioate, a phosphoramidothioate, a phosphoramidate, a phosphordiamidate, a methylphosphonate, an alkyl phosphotriester, and a formacetal, or an analog of any of these backbones.

- 39 -

In yet another embodiment, the antisense oligonucleotide is an α -anomeric oligonucleotide. An α -anomeric oligonucleotide forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gautier et al., *Nucl. Acids. Res.* 15:6625, 1987). The oligonucleotide is a 2'-O-methylribonucleotide (Inoue et al., *Nucl. Acids Res.* 15:6131, 1987), or a chimeric RNA-DNA analog (Inoue et al., *FEBS Lett.* 215:327, 1987).

Antisense oligonucleotides of the invention can be synthesized by standard methods known in the art, e.g. by use of an automated DNA synthesizer (such as are commercially available from Biosearch, Applied Biosystems, etc.). As examples, phosphorothioate oligonucleotides can be synthesized by the method of Stein et al. (*Nucl. Acids Res.* 16:3209, 1988), and methylphosphonate oligonucleotides can be prepared by use of controlled pore glass polymer supports (Sarin et al., *Proc. Natl. Acad. Sci. USA* 85:7448, 1988).

While antisense nucleotides complementary to the neurotactin coding region sequence could be used, those complementary to the transcribed untranslated region are most preferred.

One example of a 15 nucleotide antisense sequence to the human neurotactin gene is 5'-TATCGGAGCCCATGGC-3' (SEQ ID NO:5), where the underlined sequence represents the complement of the initiator methionine codon.

The antisense molecules should be delivered to cells that express neurotactin *in vivo*, e.g., brain, heart, kidney, lung, uterus, endothelial cells, fibroblasts, and bone marrow stromal cells. A number of methods have been developed for delivering antisense DNA or RNA to cells; e.g., antisense molecules can be injected directly into the tissue site, or modified antisense molecules, designed to target the desired cells

- 40 -

(e.g., antisense linked to peptides or antibodies that specifically bind receptors or antigens expressed on the target cell surface) can be administered systemically.

However, it is often difficult to achieve
5 intracellular concentrations of the antisense molecule sufficient to suppress translation of endogenous mRNAs. Therefore, a preferred approach uses a recombinant DNA construct in which the antisense oligonucleotide is placed under the control of a strong pol III or pol II
10 promoter. The use of such a construct to transfect target cells in the patient will result in the transcription of sufficient amounts of single stranded RNAs that will form complementary base pairs with the endogenous neurotactin transcripts and thereby prevent
15 translation of the neurotactin mRNA. For example, a vector can be introduced in vivo such that it is taken up by a cell and directs the transcription of an antisense RNA. Such a vector can remain episomal or become chromosomally integrated, as long as it can be
20 transcribed to produce the desired antisense RNA.

Such vectors can be constructed by recombinant DNA technology methods standard in the art. Vectors can be plasmid, viral, or others known in the art, used for replication and expression in mammalian cells.
25 Expression of the sequence encoding the antisense RNA can be by any promoter known in the art to act in mammalian, preferably human cells. Such promoters can be inducible or constitutive. Such promoters include, but are not limited to: the SV40 early promoter region (Bernoist
30 et al., *Nature* 290:304, 1981); the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto et al., *Cell* 22:787-797, 1988); the herpes thymidine kinase promoter (Wagner et al., *Proc. Natl. Acad. Sci. USA* 78:1441, 1981); or the regulatory

- 41 -

sequences of the metallothionein gene (Brinster et al., *Nature* 296:39, 1988).

Any type of plasmid, cosmid, YAC, or viral vector can be used to prepare the recombinant DNA construct
5 which can be introduced directly into the tissue site; e.g., the brain, heart, kidney, lung, uterus, endothelial cells, fibroblasts, and bone marrow stromal cells. Alternatively, viral vectors can be used that selectively infect the desired tissue (e.g., for brain, herpesvirus
10 vectors may be used), in which case administration can be accomplished by another route (e.g., systemically).

Ribozymes

Ribozyme molecules designed to catalytically cleave neurotactin mRNA transcripts also can be used to
15 prevent translation of neurotactin mRNA and expression of neurotactin (see, e.g., PCT Publication WO 90/11364; Saraver et al., *Science* 247:1222, 1990). While various ribozymes that cleave mRNA at site-specific recognition sequences can be used to destroy neurotactin mRNAs, the
20 use of hammerhead ribozymes is preferred. Hammerhead ribozymes cleave mRNAs at locations dictated by flanking regions that form complementary base pairs with the target mRNA. The sole requirement is that the target mRNA have the following sequence of two bases: 5'-UG-3'.
25 The construction and production of hammerhead ribozymes is well known in the art (Haseloff et al., *Nature* 334:585, 1988). There are numerous examples of potential hammerhead ribozyme cleavage sites within the nucleotide sequence of human neurotactin cDNA (Fig. 2). Preferably,
30 the ribozyme is engineered so that the cleavage recognition site is located near the 5' end of the neurotactin mRNA, i.e., to increase efficiency and minimize the intracellular accumulation of non-functional mRNA transcripts.

- 42 -

Examples of potential ribozyme sites in human neurotactin include 5'-UG-3' sites which correspond to the initiator methionine codon (nucleotides 87-88) and the codons for each of the cysteine residues of the chemokine-like domain (e.g., nucleotides 179-180, 191-192, 257-258, and 305-306).

The ribozymes of the present invention also include RNA endoribonucleases (hereinafter "Cech-type ribozymes"), such as the one that occurs naturally in *Tetrahymena Thermophila* (known as the IVS or L-19 IVS RNA), and which has been extensively described by Cech and his collaborators (Zaug et al., *Science* 224:574, 1984; Zaug et al., *Science* 231:470, 1986; Zug et al., *Nature* 324:429, 1986; PCT Application No. WO 88/04300; and Been et al., *Cell* 47:207, 1986). The Cech-type ribozymes have an eight base-pair sequence that hybridizes to a target RNA sequence, whereafter cleavage of the target RNA takes place. The invention encompasses those Cech-type ribozymes that target eight base-pair active site sequences present in neurotactin.

As in the antisense approach, the ribozymes can be composed of modified oligonucleotides (e.g., for improved stability, targeting, etc.), and should be delivered to cells which express the neurotactin *in vivo*, e.g., brain, heart, kidney, lung, uterus, endothelial cells, fibroblasts, and bone marrow stromal cells. A preferred method of delivery involves using a DNA construct "encoding" the ribozyme under the control of a strong constitutive *pol* III or *pol* II promoter, so that transfected cells will produce sufficient quantities of the ribozyme to destroy endogenous neurotactin messages and inhibit translation. Because ribozymes, unlike antisense molecules, are catalytic, a lower intracellular concentration is required for efficiency.

- 43 -

Other Methods for Reducing Neurotactin Expression

Endogenous neurotactin gene expression can also be reduced by inactivating or "knocking out" the neurotactin gene or its promoter using targeted homologous recombination (see, e.g., U.S. Patent No. 5,464,764). For example, a mutant, non-functional neurotactin (or a completely unrelated DNA sequence) flanked by DNA homologous to the endogenous neurotactin gene (either the coding regions or regulatory regions of the neurotactin gene) can be used, with or without a selectable marker and/or a negative selectable marker, to transfect cells that express neurotactin *in vivo*. Insertion of the DNA construct, via targeted homologous recombination, results in inactivation of the neurotactin gene. Such approaches are particularly suited for use in the agricultural field where modifications to ES (embryonic stem) cells can be used to generate animal offspring with an inactive neurotactin. However, this approach can be adapted for use in humans, provided the recombinant DNA constructs are directly administered or targeted to the required site *in vivo* using appropriate viral vectors, e.g., herpes virus vectors for delivery to brain tissue; e.g., the arcuate nucleus or the choroid plexus.

Alternatively, endogenous neurotactin gene expression can be reduced by targeting deoxyribonucleotide sequences complementary to the regulatory region of the neurotactin gene (i.e., the neurotactin promoter and/or enhancers) to form triple helical structures that prevent transcription of the neurotactin gene in target cells in the body (Helene, *Anticancer Drug Des.* 6:569, 1981; Helene et al., *Ann. N.Y. Acad. Sci.* 660:27, 1992; and Maher, *Bioassays* 14:807, 1992).

- 44 -

The Identification of Proteins which Interact with Neurotactin

The invention also features proteins which interact with neurotactin. Any method suitable for
5 detecting protein-protein interactions may be employed for identifying transmembrane proteins, intracellular, or extracellular proteins that interact with neurotactin. Among the traditional methods which may be employed are co-immunoprecipitation, crosslinking and co-purification
10 through gradients or chromatographic columns of cell lysates or proteins obtained from cell lysates and the use of neurotactin to identify proteins in the lysate that interact with the neurotactin. For these assays, the neurotactin polypeptide can be a full length
15 neurotactin, a soluble extracellular domain of neurotactin, or some other suitable neurotactin polypeptide. Once isolated, such an interacting protein can be identified and cloned and then used, in conjunction with standard techniques, to identify
20 proteins with which it interacts. For example, at least a portion of the amino acid sequence of a protein which interacts with the neurotactin can be ascertained using techniques well known to those of skill in the art, such as via the Edman degradation technique. The amino acid
25 sequence obtained may be used as a guide for the generation of oligonucleotide mixtures that can be used to screen for gene sequences encoding the interacting protein. Screening may be accomplished, for example, by standard hybridization or PCR techniques. Techniques for
30 the generation of oligonucleotide mixtures and the screening are well-known. (Ausubel, *supra*; and *PCR Protocols: A Guide to Methods and Applications*, 1990, Innis et al., eds. Academic Press, Inc., New York, NY).

Additionally, methods may be employed which result
35 directly in the identification of genes which encode proteins which interact with neurotactin. These methods

- 45 -

include, for example, screening expression libraries, in a manner similar to the well known technique of antibody probing of λ gt11 libraries, using labeled neurotactin polypeptide or a neurotactin fusion protein, e.g., an
5 neurotactin polypeptide or domain fused to a marker such as an enzyme, fluorescent dye, a luminescent protein, or to an IgFc domain.

In a related aspect, the invention features a method of identifying a compound that modulates the
10 expression or activity of neurotactin. The method is carried out by assessing the expression or activity of neurotactin in the presence and absence of the potential modulatory compound.

Methods which can be used to detect protein
15 interaction are known to those of skill in the art. For example, one method which detects protein interactions *in vivo* is the two-hybrid system (Chien et al., *Proc. Natl. Acad. Sci. USA* 88:9578, 1991). A kit for practicing this method is available from Clontech (Palo
20 Alto, CA).

Briefly, utilizing such a system, plasmids are constructed that encode two hybrid proteins: one plasmid includes a nucleotide sequence encoding the DNA-binding domain of a transcription activator protein fused to a
25 nucleotide sequence encoding neurotactin, a neurotactin polypeptide, or a neurotactin fusion protein, and the other plasmid includes a nucleotide sequence encoding the transcription activator protein's activation domain fused to a cDNA encoding an unknown protein which has been
30 recombined into this plasmid as part of a cDNA library. The DNA-binding domain fusion plasmid and the cDNA library are transformed into a strain of the yeast *Saccharomyces cerevisiae* that contains a reporter gene (e.g., HBS or *lacZ*) whose regulatory region contains the
35 transcription activator's binding site. Either hybrid

- 46 -

protein alone cannot activate transcription of the reporter gene: the DNA-binding domain hybrid cannot because it does not provide activation function and the activation domain hybrid cannot because it cannot
5 localize to the activator's binding sites. Interaction of the two hybrid proteins reconstitutes the functional activator protein and results in expression of the reporter gene, which is detected by an assay for the reporter gene product.

10 The two-hybrid system or related methodology may be used to screen activation domain libraries for proteins that interact with the "bait" gene product. By way of example, and not by way of limitation, neurotactin may be used as the bait gene product. Total genomic or
15 cDNA sequences are fused to the DNA encoding an activation domain. This library and a plasmid encoding a hybrid of bait neurotactin gene product fused to the DNA-binding domain are cotransformed into a yeast reporter strain, and the resulting transformants are screened for
20 those that express the reporter gene. For example, a bait neurotactin gene sequence, such as neurotactin or a domain of neurotactin can be cloned into a vector such that it is translationally fused to the DNA encoding the DNA-binding domain of the GAL4 protein. These colonies
25 are purified and the library plasmids responsible for reporter gene expression are isolated. DNA sequencing is then used to identify the proteins encoded by the library plasmids.

 A cDNA library of the cell line from which
30 proteins that interact with bait neurotactin gene product are to be detected can be made using methods routinely practiced in the art. According to the particular system described herein, for example, the cDNA fragments can be inserted into a vector such that they are translationally
35 fused to the transcriptional activation domain of GAL4.

- 47 -

This library can be co-transformed along with the bait neurotactin gene-GAL4 fusion plasmid into a yeast strain which contains a lacZ gene driven by a promoter which contains GAL4 activation sequence. A cDNA encoded
5 protein, fused to GAL4 transcriptional activation domain, that interacts with bait neurotactin gene product will reconstitute an active GAL4 protein and thereby drive expression of the HIS3 gene. Colonies which express HIS3 can then be purified from these strains, and used to
10 produce and isolate the bait neurotactin gene-interacting protein using techniques routinely practiced in the art.

Neurotactin and the Treatment of Inflammation

Because neurotactin is highly expressed in the brain, is up-regulated in response to inflammatory
15 stimuli (e.g., LPS and PMA) in endothelial cells (e.g., cells which line the vasculature) and is chemotactic for neutrophils (described in detail below), neurotactin may play a significant role in brain inflammation. Accumulation of neutrophils in tissues is a hallmark of
20 inflammation. Accordingly, undesirable inflammation of the brain associated with disorders such as viral encephalitis, multiple sclerosis, viral or bacterial meningitis, severe head trauma, stroke, neuro-degenerative diseases (e.g., Alzheimer's disease and Lou
25 Gehrig's disease), HIV encephalopathy, primary brain tumors (e.g., glioblastomas), Lupus associated cerebritis, and post-seizure brain injury, can be reduced by the administration of a compound that interferes with neurotactin expression or function.
30 Compounds which interfere with neurotactin function or expression can also be used to treat other undesirable inflammatory processes, e.g., atherosclerosis or respiratory infections.

- 48 -

Of course, in some circumstances, including certain phases of many of the above-described conditions, it may be desirable to enhance neurotactin function or expression, e.g., to recruit immune cells that will
5 resolve the primary infection or mediate an anti-tumor response.

Neurotactin as a Chemoprotective Agent

This invention also relates to the use of neurotactin polypeptides to protect myeloid cells, e.g.,
10 myeloid progenitor cells, and myeloid stem cells, from drugs or therapies which kill or injure actively dividing cells. Agents which protect myeloid progenitor cells and stem cells in this manner are referred to as chemoprotective agents. Such agents place myeloid
15 progenitor cells (e.g., stem cells) into a protected, slow cell-cycling state, thereby inhibiting or decreasing cell damage or death that could otherwise be caused by cell-cycle active chemotherapy drugs such as cytosine arabinoside, 5-fluorouracil, or hydroxyurea. The use of
20 chemoprotective agents permits the administration of higher doses of chemotherapeutics (or radiation) without compromising the ability of the patient to generate mature functional blood cells.

Many patients who undergo chemotherapy or
25 radiation therapy lose a substantial number of stem cells and other actively dividing myeloid progenitor cells. This loss causes the patients to become susceptible to infection and anemia. One approach for preventing neutropenia is to inhibit cell proliferation with low
30 doses of a molecule which inhibits cell cycling, thereby protecting the progenitor cells from the effects of chemotherapy and/or radiation therapy. After chemotherapy has ended, the protective treatment is also

- 49 -

stopped, which allows the progenitor cells to resume normal proliferation.

Any convenient *in vitro* or *in vivo* assay can be used to identify preferred neurotactin polypeptides or variants thereof which inhibit progenitor cell proliferation and are thus likely to be a suitable chemoprotective agent.

Suitable *in vitro* assays include those described by Gentile et al. (U.S. Patents Nos. 5,149,544 and 5,294,544). In these assays, bone marrow or spleen cells are stimulated with, e.g., CSF, in an *in vitro* system. The inhibitory activity of a candidate molecule (e.g., neurotactin) is assessed by determining the extent to which it decreases CSF-stimulated colony and cluster formation.

For example, a neurotactin polypeptide or variant can be tested as follows. LD cells are plated at a density of 5×10^5 cells in 0.3% agar culture medium with 10% FBS (Hyclone, Logan, UT) for assessment of CFU-GM. CFU-GM colonies (>40 cells/group) are stimulated by human rGM-CSF (100 U/ml) in combination with human rSLF (50 ng/ml). All colonies are tested in the absence or presence of different concentrations of a neurotactin polypeptide (or variant thereof) to determine the degree of inhibition of proliferation.

Colonies are scored after 14 days incubation at lowered (5%) O₂ tension, and 5% CO₂ in a humidified environment in an ESPEC N₂-O₂-CO₂ incubator BNP-210 (Taoi ESPEC Corp., South Plainfield, NJ). Three plates are scored per determination.

Suitable molecules are those which are effective to significantly inhibit colony formation by human bone marrow GM progenitor cells at concentrations of at least 200 ng/ml, preferably 100 ng/ml, more preferably 50 ng/ml, or even 10 ng/ml. By assaying a number of

- 50 -

neurotactin polypeptides it is possible to identify a domain of neurotactin which causes significant inhibition of proliferation.

In addition, inhibition of progenitor cell proliferation can be tested using an *in vivo* assay. A suitable murine model for assessing progenitor cell proliferation has been described by Cooper et al. (*Exp. Hematol.* 22:186, 1994). The results of this *in vivo* model, together with the *in vitro* assay results, are predictive of the efficacy of the tested molecules in treating patients, e.g., humans.

In suitable *in vivo* tests, molecules are evaluated for effects on myelopoiesis in mice, with endpoints being nucleated cellularity and differentials in the bone marrow, spleen, and peripheral blood, and absolute numbers and cycling status of myeloid progenitor cells in the marrow and spleen. In each test, groups of C3H/HeJ mice are exposed to a particular test sample. C3H/HeJ mice are preferred because they are relatively insensitive to the effects of endotoxin. Thus, any potential endotoxin contamination in the test samples will not influence the *in vivo* results.

Neurotactin polypeptides can be tested as follows, although other assays are also useful. C3H/HeJ mice are obtained from the Jackson Laboratory (Bar Harbor, ME) and housed in a conventional animal facility. The mice are injected intravenously with 0.2 ml/mouse sterile pyrogen-free saline, or the stated amount of a selected neurotactin polypeptide or variant as described in Mantel et al. (*Proc. Natl Acad. Sci. USA* 90:2232, 1993). The mice are sacrificed 24 hours later.

The cycling status of hematopoietic progenitor cells, i.e., the proportion of progenitor cells in DNA synthesis (S phase of the cell cycle), is estimated as described in Maze et al. (*J. Immunol.* 149:1004, 1992) and

- 51 -

Cooper et al. (*Exp. Hematol.* 22:186, 1994). The high specific activity (20 Ci/mM)-tritiated thymidine (50 μ Ci/mL) (New England Nuclear; Boston, MA) kill technique is used, and is based on a calculation *in vitro* of the reduction in the number of colonies formed after pulse exposure of cells for 20 minutes to "hot" tritiated thymidine as compared with a control such as McCoy's medium or a comparable amount of non-radioactive "cold" thymidine.

10 Femoral bone marrow is removed from the sacrificed mice, treated with high-specific-activity tritiated thymidine, and plated in 0.3% agar culture medium with 10% FBS in the presence of 10% volume/volume pokeweed mitogen mouse spleen cell cultured medium. Colonies 15 (>40 cells/aggregate) and clusters (3-40 cells) are scored after 7 days of incubation.

Three plates are scored for each sample for a statistical analysis. Each mouse is evaluated separately in groups of three mice each.

20 Preferred neurotactin polypeptides and variants are effective at a dosage of 200 μ g/mouse, 100 μ g/mouse, 50 μ g/mouse, or even 10 μ g/mouse or lower. An effective dosage will reduce progenitor cell cycling by at least 25% or at least 50% or even more.

25 Chemoprotective neurotactin polypeptides can be administered to a patient as adjunctive agents before and/or during chemotherapy or radiation therapy to protect progenitor cells from the cytotoxic effects of the chemotherapeutic agents or radiation.

30 Chemoprotective neurotactin polypeptides place myeloid cells into a protected, slow-cycling state, thereby inhibiting or decreasing cell damage that could otherwise be caused by cell-cycle active chemotherapy drugs such as cytosine arabinoside, 5-fluorouracil, or hydroxyurea.

35 The use of chemoprotective agents permits the

- 52 -

administration of higher doses of chemotherapeutics without compromising the ability of the patient to generate mature functional blood cells.

Chemoprotective neurotactin polypeptides are
5 administered to a patient in the same manner as chemokines generally, e.g., injected intravenously or subcutaneously, in a pharmaceutically acceptable carrier.

In chemotherapy, specific protocols may vary, and factors such as tumor size, growth rate, and location of
10 the tumor all affect the course of therapy. Administration of chemotherapeutic agents as well as chemoprotective agents require may required knowledge of the extent of disease, the toxicity of previous treatment courses, and the degree of the expected chemotherapeutic
15 drug toxicity.

Examples

Example 1 describes the identification and sequencing of a murine neurotactin gene and a human neurotactin gene. Example 2 describes the
20 characterization of neurotactin, including its expression pattern and its ability to act as a chemoattractant for neutrophils. Example 3 describes the chromosomal mapping of the neurotactin gene.

Example 1: Cloning of the Neurotactin Gene

25 The gene for murine neurotactin was identified in a murine choroid plexus cDNA library. This murine neurotactin gene was used to identify a human neurotactin gene. The identification and sequencing of both genes is described in this first example.

30 Choroid-Plexus mRNA Isolation: The murine mRNA used to create the murine choroid plexus library was prepared as follows. Total RNA was isolated from mouse

- 53 -

choroid plexus tissue using the guanidinium isothiocyanate/CsCl method of Chirgwin et al. (*Biochemistry* 18:5294, 1979) as described in Current Protocols for Molecular Biology (*supra*). The RNA was
5 quantitated, diluted to 1 mg/ml in water, and then incubated for 30 minutes at 37°C with an equal volume of DNase solution (20 mM MgCl₂, 2 mM DTT, 0.1 units DNase, 0.6 units RNase inhibitor in TE) to remove contaminating DNA. The RNA was then extracted with phenol-chloroform-
10 isoamyl alcohol, and ethanol precipitated. After quantitation at 260 nm, an aliquot was electrophoresed to check the integrity of the RNA. Next, PolyA⁺ RNA was isolated using an Oligotex-dT kit from Qiagen (Chatsworth, CA) as described by the manufacturer. After
15 quantitation, the mRNA was precipitated in ethanol and resuspended at a concentration of 1 mg/ml in water.

cdNA Library Construction: The isolated choroid plexus mRNA described above was used to prepare cdNA as follows.

20 Choroid plexus mRNA was used as a template for preparation of cdNA according to the method of Gubler et al. (*Gene* 25:263, 1983) using a Superscript Plasmid cdNA synthesis kit (Life Technologies; Gaithersburg, MD). The cdNA obtained was ligated into the *NotI*/*SalI* sites of
25 the mammalian expression vector pMET7, a modified version of pME18S, which utilizes the SRA promoter as described previously (Takebe, *Mol. Cell. Bio.* 8:466, 1988). Ligated cdNA was transformed into electrocompetent DH10B *E. coli* either prepared by standard procedures or
30 obtained from Life Technologies.

DNA Preparation and Sequence Analysis: A number of cdNA clones in the murine choroid plexus library were sequenced to identify sequences of interest. The

- 54 -

identified sequences were then used to clone and sequence a complete murine neurotactin gene. The identification and analysis was performed as follows.

First, 96-well plates were inoculated with
5 individual choroid plexus library transformants in 1 ml of LB-amp. These inoculations were based on the titers of the cDNA transformants. The resulting cultures were grown for 15 to 16 hours at 37°C with aeration. Prior to
10 DNA preparation, 100 µl of cell suspension was removed and added to 100 µl of 50% glycerol, mixed and stored at -80°C (glycerol freeze plate). DNA was then prepared using the Wizard™ miniprep system (Promega; Madison, WI) employing modifications for a 96-well format.

The insert cDNAs of a number of clones were
15 sequenced by standard, automated fluorescent dideoxynucleotide sequencing using dye-primer chemistry (Applied Biosystems, Inc., Foster City, CA) on Applied Biosystems 373 and 377 sequenators (Applied Biosystems). The primer used in this sequencing was proximal to the
20 SRa promoter of the vector and therefore selective for the 5' end of the clones, although other primers with this selectivity can also be used. The short cDNA sequences obtained in this manner were screened as follows.

25 First, each sequence was checked to determine if it was a bacterial, ribosomal, or mitochondrial contaminant. Such sequences were excluded from the subsequent analysis. Second, sequence artifacts, such as vector and repetitive elements, were masked and/or
30 removed from each sequence. Third, the remaining sequences were searched against a copy of the GenBank nucleotide database using the BLASTN program (BLASTN 1.3MP: Altschul et al., *J. Mol. Bio.* 215:403, 1990). Fourth, the sequences were analyzed against a non-
35 redundant protein database with the BLASTX program

- 55 -

(BLASTX 1.3MP: Altschul et al., supra). This protein database is a combination of the Swiss-Prot, PIR, and NCBI GenPept protein databases. The BLASTX program was run using the default BLOSUM-62 substitution matrix with the filter parameter: "xnu+seg". The score cutoff utilized was 75.

Assembly of overlapping clones into contigs was done using the program Sequencher (Gene Codes Corp.; Ann Arbor, MI). The assembled contigs were analyzed using the programs in the GCG package (Genetic Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711).

The above-described analysis resulted in the identification of a clone (clone ID jfmjd006h11) having an open reading frame of 395 amino acids (Fig. 1). The protein encoded by this clone was named neurotactin. The first approximately 21 amino acids in this open reading frame were predicted to be a signal sequence using the method of Von Heijne (*J. Membrane Biol.* 115:195, 1990). The amino-terminal portion of murine neurotactin has significant homology to the known C-C family of chemokines. This portion is 40% identical to mouse monocyte MCP-1 based on a primary sequence alignment of residues 23 to 92 of murine neurotactin with murine MCP-1.

The sequence of the murine neurotactin gene was used to search the DBEST expressed sequence tag (EST) database using the BLASTN program. The clone corresponding to a selected EST was obtained from Genome Systems Inc. and sequenced in full (Fig. 2). The sequence of this clone differed from that of the initially identified EST at position 319. Compared to the sequence of the clone, there is a one base pair deletion in the EST sequence which results in a reading frame shift. Because of this deletion, the predicted

- 56 -

amino acid sequence of the clone differs from the predicted amino acid sequence corresponding to the EST.

This form of human neurotactin is 42% identical to human MCP-1 based on a primary sequence alignment of
5 residue 23 to 92 of human neurotactin with human MCP-1 (Swiss Prot # 13500). Overall human neurotactin contains 397 amino acids and is 67% identical to the form of murine neurotactin described herein (Fig. 4). Higher than average homology is observed at the chemokine-like
10 region at the N-terminal. The regions having the greatest homology are the transmembrane domain and the cytoplasmic domain.

Example 2: Characterization of Neurotactin

The expression pattern of neurotactin was examined
15 as described below. Also described below is the expression of a recombinant form of soluble murine neurotactin and experiments demonstrating that neurotactin stimulates chemotaxis of neutrophils.

Analysis of Neurotactin Expression: Northern
20 analysis was used to examine neurotactin expression as follows. First, total RNA from the following cell types was extracted with RNAsol when the cells were at 80% confluence: WEH1-3 and Pu5-1.8 (myelomonocytes), P388D1 and IC-21 (macrophages), AKR.G.2 (thymoma), BaF3 (Pro B
25 cell), EL-4 (Lymphoma), NFS-1.0 (B cell lymphoma), BCL (B cell leukemia), STO (embryonic fibroblasts), EOMA (endothelial), and BMS-12 (bone marrow stromal). Except for EOMA cells and BMS-12 cells, all of these cells were maintained according to procedures described by the
30 American Type Culture Collection (Bethesda, MD). EOMA cells were maintained in DMEM with 10% FCS, and BMS-12 cells were maintained in DMEM with 10% horse serum. In order to determine the effect of activators on

- 57 -

neurotactin, cells were treated with 100 ng/ml LPS or 30 ng/ml PMA for 4 hours prior to RNA extraction or were left untreated.

Northern blots containing 20 µg of total RNA were probed using standard techniques (Chirgwin et al., *Biochemistry* 18:5294, 1979) with a ³²P-labeled DNA fragment encoding the full-length neurotactin.

This Northern analysis revealed that a 3.5 kb neurotactin mRNA is constitutively expressed in endothelial cells (EOMA) and embryonic fibroblasts (STO). The 3.5 kb mRNA is upregulated in these cells when they are stimulated with LPS and PMA. This mRNA is also upregulated in bone marrow stromal cells (BMS-12) when treated with LPS and PMA. It is not expressed by unstimulated BMS-12 cells. This expression pattern is characteristic of modulators of inflammation.

Neurotactin mRNA was not detected in several cell lines of hematopoietic origin with or without stimulation by PMA and LPS. The tested cell lines were WEHI-3 and Pu5-1.8 (myelomonocytic), P388D1 and IC-21 (monocytic/macrophage), ARK.G.2 (thymoma), BaF3 (pro B cell), EL-4 (lymphoma), NFS-1.0 (B cell lymphoma) and BCL (B cell leukemia).

This same Northern analysis revealed the presence of messages having the following approximate sizes: 1.5 kb, 4.4 kb, and 5.5 kb. These messages are likely to represent alternatively spliced forms of neurotactin or the transcription products of related genes.

Human tissue Northern blots showed that human neurotactin is highly expressed in the brain and the heart. Furthermore, hybridization to a brain tissue Northern blot (Clontech) showed that the mRNA was expressed in all parts of the brain.

In situ hybridizations were also used to examine neurotactin expression. Tissues for these hybridization

- 58 -

were prepared as follows. Four to six week old C57BL/6 mice were anesthetized and perfused with PBS followed by 4% paraformaldehyde (PFA/PBS). The brains were then removed and stored in 10% buffered formalin. Ten μ m coronal frozen sections of brain were post-fixed with 4% PFA/PBS for 15 minutes. After washing with PBS, sections were digested with 2 μ g/ml proteinase K at 37° for 15 minutes, and then incubated with 4% PFA/PBS for 10 minutes. Sections were then washed with PBS, incubated with 0.2 N HCl for 10 minutes, washed with PBS, incubated with 0.25% acetic anhydride/1 M triethanolamine for 10 minutes, washed with PBS, and dehydrated with 70% ethanol and 100% ethanol.

Hybridizations were performed with 35 S-radiolabeled (5×10^7 cpm/ml) antisense cRNA probes encoding a 1.9 kb segment of the coding region of the murine neurotactin gene (clone 6h11) in the presence of 50% formamide, 10% dextran sulfate, 1X Denhardt's solution, 600 mM NaCl, 10 mM DTT, 0.25% SDS, and 100 μ g/ml tRNA for 18 hours at 55°C. After hybridization, the slides were washed with 5X SSC at 55°C, 50% formamide/2X SSC at 55°C for 30 minutes, 10 mM Tris-HCl (pH 7.6)/500 mM NaCl/1 mM EDTA (TNE) at 37°C for 10 minutes, incubated in 10 μ g/ml RNase A in TNE at 37°C for 30 minutes, washed in TNE at 37°C for 10 minutes, incubated once in 2X SSC at 50°C for 30 minutes, twice in 0.2X SSC at 50° C for 30 minutes, and dehydrated with 70% ethanol and 100% ethanol. Localization of mRNA transcripts was detected by dipping slides in Kodak NBT-2 photoemulsion and exposed for 4 days at 4°C. Controls for the *in situ* hybridization experiments included the use of a sense probe which showed no signal above background levels.

These *in situ* hybridizations with murine neurotactin probe showed that neurotactin is expressed in

- 59 -

the arcuate nucleus. Expression was also detected in the cortex and choroid plexus.

To determine the association of neurotactin with the cell membrane, a construct containing the full-length
5 neurotactin coding region was made in a mammalian expression vector, pN83. Full-length murine neurotactin cDNA was modified for expression in a mammalian system by PCR with the following primers:
5'-GGGAAAGAATTCATGGCTCCCTCGCCGCTCGCGTCC-3' (SEQ ID NO:12)
10 and
5'-GGGAAACTCGAGTCATTTATCATCATCATCTTTATAATCCACTGGCACCAGGACGTATGA-3' (SEQ ID NO:13). Nucleotides encoding a FLAG epitope tag (DYKDDDDK (SEQ ID NO:14), which can be detected with an M2 anti-FLAG antibody, were incorporated
15 into the 3' "reverse" primer. The PCR products were cloned into the pN83 vector bearing the EBV origin of replication. The construct DNA was prepared with the Qiagen Maxiprep™ kit (Qiagen, Chatsworthy, CA) and transfected with lipofectamine™ (Gibco, Gaithersburg, MD)
20 into 293 EBNA cells that were cultured in 8-well chamber slides. Forty-eight hours after transfection, the cells were fixed with 50% methanol and 50% acetone for 1 minute at room temperature, washed four times with 2.5 ml TBS, and incubated with 10 µg/ml of M2 anti-FLAG monoclonal
25 antibody (Eastman Kodak Co., New Haven, CT) and then with FITC-conjugated goat anti-mouse antibody at 1:1000 dilution (Jackson Immuno Research, West Grove, PA). The cells were exposed to the primary and secondary antibodies for one hour each. The immunofluorescent
30 staining was visualized under 200-fold magnification. Strong staining was detected on the surface of the transfected 293 EBNA cells, suggesting full-length neurotactin is indeed membrane-anchored. The signal observed could be competed out by adding an excess of the
35 FLAG peptide to the incubation.

- 60 -

Preparation of Soluble Neurotactin A soluble form of recombinant murine neurotactin (residues 22-105) was produced in bacteria using the pGEX expression system. The pGEX-neurotactin was purified on glutathione agarose and the neurotactin moiety released by thrombin digestion. Following endotoxin removal on an Endotoxin BX column (Cape Cod Associates: Falmouth, MA) the neurotactin preparation was determined to contain low levels of endotoxin (<0.01 EU/ml) by the Limulus
10 ameocyte lysate (LAL) assay (Cape Cod Associates).

Recombinant, soluble neurotactin was produced as follows. First, the coding region of mouse neurotactin was amplified with a primer corresponding to a sequence at the 5' end of the sequence encoding the chemokine-like
15 domain (5' primer). The 5' primer, 5'-GGGAAAGAATTCCTGCCGGGTCAGCACCTCGGCATG-3' (SEQ ID NO:6), has an *EcoRI* restriction enzyme cleavage site followed by 24 nucleotides encoding the beginning of the chemokine-like domain of murine neurotactin. The 3' primer used
20 was 5'-GGGAAACTCGAGTCATTCTCAAACCTTGCCACCATTTTTTA-3' (SEQ ID NO:7). This primer has complementary sequences encoding amino acids 99 to 105 preceded by a termination codon and *XhoI* site.

These primers pairs were used for PCR
25 amplification using the following conditions: 94°C for 30 seconds; 55°C for 30 seconds and 72°C for 90 seconds with 30 cycles. The resulting PCR product was cloned into the GST fusion protein vector pGEX-4T (Pharmacia, Piscataway, NJ). The fusion protein was produced in
30 *E. coli* and purified according to the protocol supplied by the manufacturer. The neurotactin construct produced a protein of approximately 10,000 kDa after the cleavage of GST by thrombin.

- 61 -

Assay of Neurotactin as a Neutrophil

Chemoattractant The ability of a soluble form of recombinant neurotactin described above was tested for its ability to act as a chemoattractant for neutrophils.

- 5 The human neutrophils used in these experiments were isolated by layering 20 ml of human blood over 10 ml of FICOLL 1119 and 10 ml of FICOLL 1077 (Sigma; St Louis, MO) in a 50 ml conical polypropylene tube and centrifuging at 1800 rpm for 15 minutes at room
- 10 temperature. Following centrifugation, the neutrophil layer was washed with sterile, ice cold, calcium and magnesium-free phosphate buffered saline (Gibco; Bethesda, MD), and the neutrophil pellet was resuspended in 45 ml sterile, cold H₂O and 5 ml sterile 10X phosphate
- 15 buffered saline (PBS) to lyse contaminating red blood cells. The cells were pelleted by centrifugation at 1800 rpm for 5 minutes at 4°C. The pelleted neutrophils were resuspended in 10 ml of sterile PBS, quantitated, and kept on ice until needed for *in vitro* assays.
- 20 The ability of neurotactin to elicit neutrophil chemotaxis was tested in an *in vitro* assay as described by Falk et al. (*J. Immunol. Meth.* 33:39, 1980). Approximately 50,000 neutrophils were added to each well of a 48-well micro chemotaxis chamber containing a 5 μ m
- 25 pore size filter. Varying concentrations of neurotactin or human interleukin-8 (Biosource International, Camarillo, CA) were added to the lower chamber of each appropriate well. The cells were then incubated at 37°C, 5% CO₂ for 30 minutes. The upper chamber was removed and
- 30 the cells on the filter from the upper chamber were removed by scraping. The filter was fixed in 100% ethanol and stained with a solution of 0.5% toluidine blue in 3.7% formaldehyde. Excess stain was removed with distilled water. Migrated cells were quantitated by
- 35 counting three high power fields (400X magnification) per

- 62 -

well. The number of cells that migrated to the buffer control were subtracted as background. The activities of compounds in this assay are predictive of the ability of compounds to elicit neutrophil chemotaxis *in vivo*.

5 The results of this assay are presented in Fig. 5. This figure provides a comparison of the ability of human interleukin-8 and neurotactin to elicit neutrophil chemotaxis. As can be seen in the figure, neurotactin is chemotactic for human neutrophils with an approximate
10 endpoint titer (concentration giving 50% of the maximal stimulation level) of 1 ng/ml. This endpoint titer compares favorably with that of human interleukin-8, a known neutrophil chemoattractant.

A soluble version of murine neurotactin,
15 corresponding to the chemokine-like domain (residues 22 to 105), was also chemotactic for human T lymphocytes. Further testing showed that neither this chemokine-like domain nor an entire extracellular domain had a chemotactic effect on human monocytes, human
20 myelomonocytic THP1 cells, or murine P388D1 monocytic cells.

Two soluble versions of murine neurotactin (corresponding to the chemokine-like domain (residues 22 to 105) and the entire extracellular domain (residues 22
25 to 337)) were also injected into C57BL/6J mice intraperitoneally to determine the chemotactic efficacy *in vivo*. Bacterial lysate containing only glutathione S-transferase was purified in the same fashion as these soluble forms of neurotactin and served as a control for
30 the *in vivo* assay. Neurotactin and the control protein were both administered at a dose of 0.5 μ g/400 μ l PBS/mouse. In addition, some mice received only 400 μ l of PBS. Two hours after the injection, peritoneal exudate was collected and the number and subtype of
35 leukocytes recovered in the fluid were determined by

- 63 -

typing and counting the cells in four high power fields (40X magnification; total area 0.5 mm²). The neutrophils and eosinophils were identified by staining with Wright-Giemsa, and lymphocytes were assessed by Thy 1.2 (53-2.1) 5 and IgM (II/41) immunostaining.

The chemokine-like domain (residues 22 to 105) was chemotactic for neutrophils and lymphocytes (as predicted from *in vitro* studies), and was also chemotactic for monocytes. However, the activity of the chemokine-like 10 domain towards monocytes may be due to a secondary effect *in vivo* and may not, therefore, reflect the chemotactic specificity of neurotactin. The chemokine-like domain of neurotactin was not chemotactic for eosinophils. The entire extracellular domain (residues 22 to 337) failed 15 to act as a chemoattractant for monocytes and lymphocytes *in vivo* but was chemotactic for neutrophils.

Neurotactin expression was determined by immunohistochemistry in normal and LPS-treated murine brain and in brain sections from mice with severe 20 experimental autoimmune encephalomyelitis (EAE). EAE develops spontaneously in immunodeficient α -myelin basic protein T cell receptor transgenic mice, which are used as a model of chronic inflammatory diseases.

Polyclonal anti-neurotactin antibody was raised in 25 rabbits against a peptide located at the amino terminus of neurotactin (LPGQHLGMTKCEIM (SEQ ID NO:15); Research Genetics, Huntsville, AL). The antibody was affinity purified from 12-week bleeds. For LPS treatment, 8 week old CD1 mice were injected intravenously with 40 μ g LPS 30 and sacrificed 2 hours later by cervical dislocation. Their brains were removed, bisected transversely, and coated with Tissue Tek[™] OCT compound (Cryoform). The OCT-coated tissue was then snap frozen in a mixture of isopentane and dry ice and stored at -70°C. Sections 35 (3 μ m thick) were cut onto microscope slides, air dried,

- 64 -

fixed in 2% paraformaldehyde (for 5 minutes at 4°C) and methanol (10 minutes at -20°C). To study neurotactin expression in the EAE model, brains were collected from male α -myelin basic protein T cell receptor (MBP TCR) transgenic mice (at about 4 months of age) and fixed in 10% neutral buffered formalin and embedded in paraffin. Sections (4 μ m) were microwaved twice for 5 minutes in 0.01 M sodium citrate (pH 6.0) before staining.

The fixed sections were stained with antibody using an avidin/biotin staining method. All incubations were carried out under humidified conditions and slides were washed twice between the steps for 5 minutes each in 0.1 M phosphate buffered saline supplemented with 0.2% gelatin (PBSG). The sections were overlaid with 20% fetal calf serum in PBS for 15 minutes and then incubated overnight at 4°C with polyclonal anti-neurotactin or normal rabbit serum (both diluted to 1:200 in PBS supplemented with 0.1% bovine serum albumin). Endogenous peroxide was blocked by incubation for 20 minutes in methanol containing 0.3% hydrogen peroxide. Non-specific staining due to cross reaction with endogenous avidin or biotin was blocked by incubation with avidin solution followed by biotin solution, both for 20 minutes. Bound monoclonal antibody was visualized by incubation with biotinylated swine anti-rabbit immunoglobulin (Dako, CA) and then streptavidin peroxidase complex, both diluted in 10% normal mouse serum with PBS, and incubated for one hour. The slides were then flooded with peroxidase substrate solution (400 μ g diaminobenzidine in 10 ml PBS containing 0.01% hydrogen peroxide) for 10 minutes. The sections were counterstained with haematoxylin. Control sections were generated by selectively omitting monoclonal antibody, biotinylated anti-rat immunoglobulin or streptavidin complex. In addition, competitive

- 65 -

inhibition of the antibody was accomplished by preincubation of antibody with the peptide (25 µg/ml) for 45 minutes at 37°C prior to incubation with the tissue sections.

5 In the normal mouse brain, staining was localized to capillary vessels and resident microglia. An increased intensity of labeling was observed on the same cell types two hours after LPS treatment. In addition, an increased number of activated microglial cells stained
10 positive for the anti-neurotactin antibody. In both normal and LPS-treated brain, staining of larger vessels was restricted to the apical region of the endothelium. It is known that, in addition to resident microglial cells, two other subtypes of microglial cell are present
15 in the CNS at the blood-brain barrier: perivascular and juxtavascular microglia. The anti-neurotactin antibody staining associated with micro vessels was consistent with the staining of those microglial cells. Neurotactin expression was also up-regulated in activated microglial
20 in the brains of EAE mice.

Example 3: Mapping of the Neurotactin Gene

Described in this example is the chromosome mapping of neurotactin. Also described below is the potential relationship between neurotactin and Bardet-
25 Biedl Syndrome.

Mouse Chromosome Mapping: The following PCR primers were used to amplify mouse genomic DNA.

Forward primer: 5'-CACAGTCCACCCCTCAG-3' (SEQ ID NO:8)

30 Reverse primer, 5'-GCTCTGGTAAGCAAACATGG-3' (SEQ ID NO:9).

PCR reactions were performed on genomic DNA from a panel of interspecific backcrossed mice. The

- 66 -

amplification profile was as follows: 94°C (30 seconds); 55°C (30 seconds); and 72°C (45 seconds) with 30 cycles. Samples were run on nondenaturing 10% acrylamide SSCP gel at 20 W and 4°C for 2.5 hours.

5 Murine neurotactin was mapped to the long arm of mouse chromosome 8, between D8MIT35 and D8MIT74, by using a panel of backcrossed progeny of the C57Bl/6J and *Mus spretus*. The region is syntenic to human chromosome 16.

10 Human Chromosome Mapping: Human neurotactin was mapped to chromosome 16q, between W17078 and WI6174, using a panel of radiation hybrids.

The following primers were used to amplify human genomic DNA from a panel of radiation hybrids (Genebridge
15 4, Research Genetics, Huntsville, AL).

Forward: 5'-TGTGAACTCCTCTGGCCTGT-3' (SEQ ID
NO:10)

Reverse: 5'-GAAGGGGCTGGGCATTTAAT-3' (SEQ ID
NO:11)

20 The amplification profile was as follows: 94°C for 30 seconds; 55°C for 30 seconds, and 72°C for 45 seconds with 30 cycles. Samples were resolved on 1% agarose TAE gel.

Based on a published article (Kwitek-Black et al.,
25 *Nature Genetics* 5:392, 1993) and the integrated genetic map of Chromosome 16 (Genome Directory, *Nature* 377:335, 1995), the region to which neurotactin gene maps overlaps with a locus for the a gene important in Bardet-Biedl Syndrome (BBS).

30 BBS is a heterogeneous autosomal recessive disorder characterized by obesity, mental retardation, polydactyly, retinitis pigmentosa and hypogonadism. Patients suffering from this syndrome have a high incident of renal and cardiovascular abnormalities. The

- 67 -

fact that neurotactin is expressed in tissues and organs which may be affected in BBS suggests that neurotactin may play a role in BBS. For example, BBS is characterized by obesity and mental retardation, and
5 neurotactin is expressed in the arcuate nucleus, a region of the brain thought to play a role in weight control, and other parts of the brain. BBS is also associated with renal and cardiovascular symptoms, and neurotactin is expressed in the kidney and heart. In addition, BBS
10 is associated with hypogonadism, and neurotactin is expressed in the uterus.

Therapeutic Applications

The neurotactin proteins and polypeptides described herein stimulate chemotaxis of neutrophils are
15 thus likely to mediate inflammation. Accordingly, undesirable inflammation of the brain associated with disorders such as viral encephalitis, multiple sclerosis, viral or bacterial meningitis, severe head trauma, stroke, neuro-degenerative diseases (e.g., Alzheimer's
20 disease and Lou Gehrig's disease), HIV encephalopathy, primary brain tumors (e.g., glioblastomas), Lupus associated cerebritis, and post-seizure brain injury, can be reduced by the administration of a compound that interferes with neurotactin expression or function (e.g.,
25 an antibody). Compounds which interfere with neurotactin function may also be used to treat other undesirable inflammatory processes, e.g., atherosclerosis or respiratory infections.

Neurotactin, like other chemokines (Lord et al.,
30 *Blood* 85:3412, 1995; Laterveer et al., *Blood* 85:2269, 1995), can be used to mobilize hematopoietic stem cells and progenitor cells from the bone marrow to the peripheral blood. Because stem cells and progenitor cells can be more easily recovered from the peripheral

- 68 -

blood than from bone marrow, neurotactin may be useful for isolating such cells for use in stem cell restorative therapy. Such therapy is useful for patients which have undergone myeloablative and/or myelosuppressive cancer
5 treatments.

Neurotactin is likely to be involved in the regulation of hematopoietic cells. In particular, neurotactin, like other chemokines (Graham et al., *Nature* 344:442, 1994; Broxmeyer et al., *J. Immunol.* 150:3448,
10 1993), may be able to inhibit proliferation of hematopoietic stem cells and progenitor cells. Such inhibition can protect the cells from chemotherapeutic damage. Thus, neurotactin can be used to protect hematopoietic stem cells and progenitor cells from
15 chemotherapeutic damage, e.g, damage during chemotherapy for cancer.

The neurotactin polypeptides which inhibit progenitor cell proliferation can be used to inhibit hyperproliferative myeloid based diseases such as chronic
20 myelogenous leukemia, polycythemia vera, and hypermegakaryocytopoietic disorders. Hyperproliferative states in such disorders occur because the progenitor cells are unable to negatively regulate cell growth and replication. Administration of suitable neurotactin
25 polypeptides is expected to inhibit cell replication resulting in the inhibition of the abnormal cell growth. Dosages of the neurotactin polypeptides for treating hyperproliferative myeloid based diseases would be similar to those dosages described above for use of the
30 proteins as adjuncts to chemotherapy.

In addition, neurotactin polypeptides can be used to prevent myeloid progenitor cells from becoming leukemic as the result of the administration of chemotherapeutic agents. The neurotactin polypeptides are
35 administered in the same way described above.

- 69 -

Recombinant neurotactin may facilitate the production of pharmacologic modifiers of neurotactin function. Such therapeutic polypeptides of the invention can be administered by any appropriate route, e.g.,
5 intravenously, at a dosage that is effective to modulate neurotactin function. Treatment may be repeated as necessary for alleviation of disease symptoms.

Diagnostic Applications

The nucleic acids, polypeptides, and antibodies of
10 the invention are useful for identifying those compartments of mammalian cells which contain proteins important to the function of neurotactin. Antibodies specific for neurotactin may be produced as described above. The normal subcellular location of the protein is
15 then determined either *in situ* or using fractionated cells by any standard immunological or immunohistochemical procedure (see, e.g., Ausubel et al., *supra*; Bancroft and Stevens, *Theory and Practice of Histological Techniques*, Churchill Livingstone, 1982).
20 Antibodies specific for neurotactin also find diagnostic use in the detection or monitoring of neurotactin-related diseases. Levels of a neurotactin protein in a sample can be assayed by any standard technique. For example, neurotactin protein expression
25 can be monitored by standard immunological or immunohistochemical procedures (e.g., those described above) using the antibodies described herein. Alternatively, neurotactin expression can be assayed by standard Northern blot analysis or can be aided by PCR
30 (see, e.g., Ausubel et al., *supra*; *PCR Technology: Principles and Applications for DNA Amplification*, ed., H.A. Ehrlich, Stockton Press, NY). If desired or necessary, analysis can be carried out to detect point mutations in the neurotactin sequence (for example, using

- 70 -

well known nucleic acid mismatch detection techniques). All of the above techniques are enabled by the neurotactin sequences described herein.

Accordingly, the nucleic acids, polypeptides and
5 antibodies of the invention can be used in a method for determined whether a patient has a disorder associated with abnormal expression of neurotactin. The method can be carried out by quantitating the level of expression of neurotactin in a biological sample obtained from the
10 patient. As a control, the quantitation can be carried out using a biological sample obtained from a patient who is healthy.

Neurotactin expression can be assessed at the level of gene expression, for example, by quantitating
15 the level of neurotactin mRNA expression in the biological sample, or at the level of protein expression, by quantitating the level of neurotactin protein expressed. Quantitation can be carried out using the techniques described above, which are well within the
20 abilities of those of skill in the art to perform.

Should it be determined that a patient has a disorder that is associated with abnormal expression or activity of neurotactin, the patient can be given a compound that modulates that expression or activity. For
25 example, the patient can receive a compound such as a small molecule, an antisense nucleic acid molecule, or a ribozyme, that inhibits the expression of neurotactin. The patient can also receive a compound that inhibits the activity of neurotactin. An antibody that specifically
30 binds neurotactin can be used for this purpose. Alternatively, the patient can receive a compound that enhances the expression or activity of neurotactin. Compounds that inhibit or enhance the expression or activity of neurotactin can include synthetic molecules.
35 These methods of treatment can be used to treat

- 71 -

inflammatory disorders and disorders associated with cellular proliferation, as described more fully above.

Other Embodiments

The invention also features fragments, variants, 5 analogs and derivatives of the neurotactin polypeptides described above that retain one or more of the biological activities of neurotactin such as neutrophil chemotaxis.

The invention includes naturally-occurring and non-naturally-occurring allelic variants. Compared to 10 the most common naturally-occurring nucleotide sequence encoding neurotactin, the nucleic acid sequence encoding allelic variants may have a substitution, deletion, or addition of one or more nucleotides. The preferred allelic variants are functionally equivalent to 15 naturally-occurring neurotactin.

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, that the foregoing description is intended to illustrate and not limit the scope of the 20 invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

- 72 -

SEQUENCE LISTING

(1) GENERAL INFORMATION

- (i) APPLICANT: Millennium Pharmaceuticals, Inc.
- (ii) TITLE OF THE INVENTION: NEUROTACTIN AND USES THEREFOR
- (iii) NUMBER OF SEQUENCES: 15
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Fish & Richardson, P.C.
 - (B) STREET: 225 Franklin Street
 - (C) CITY: Boston
 - (D) STATE: MA
 - (E) COUNTRY: US
 - (F) ZIP: 02110-2804
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: Windows95
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: PCT/US97/----
 - (B) FILING DATE: 06-MAY-1997
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 08/643,798
 - (B) FILING DATE: 07-MAY-1996
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Fasse, J. Peter
 - (B) REGISTRATION NUMBER: 32,983
 - (C) REFERENCE/DOCKET NUMBER: 07334/029WO1
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 617-542-5070
 - (B) TELEFAX: 617-542-8906

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1784 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 40...1224

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GTGACCCAC GCGTCCGGCC GAATTCCTGC ACTCCAGCC ATG GCT CCC TCG CCG
Met Ala Pro Ser Pro
1 5

54

- 73 -

CTC GCG TGG CTG CTG CGC CTG GCC GCG TTC TTC CAT TTG TGT ACT CTG Leu Ala Trp Leu Leu Arg Leu Ala Ala Phe Phe His Leu Cys Thr Leu	102
10 15 20	
CTG CCG GGT CAG CAC CTC GGC ATG ACG AAA TGC GAA ATC ATG TGC GAC Leu Pro Gly Gln His Leu Gly Met Thr Lys Cys Glu Ile Met Cys Asp	150
25 30 35	
AAG ATG ACC TCA CGA ATC CCA GTG GCT TTG CTC ATC CGC TAT CAG CTA Lys Met Thr Ser Arg Ile Pro Val Ala Leu Leu Ile Arg Tyr Gln Leu	198
40 45 50	
AAC CAG GAG TCC TGC GGC AAG CGT GCC ATT GTC CTG GAG ACG ACA CAG Asn Gln Glu Ser Cys Gly Lys Arg Ala Ile Val Leu Glu Thr Thr Gln	246
55 60 65	
CAC AGA CGC TTC TGT GCT GAC CCG AAG GAG AAA TGG GTC CAA GAC GCC His Arg Arg Phe Cys Ala Asp Pro Lys Glu Lys Trp Val Gln Asp Ala	294
70 75 80 85	
ATG AAG CAT CTG GAT CAC CAG GCT GCT GCC CTC ACT AAA AAT GGT GGC Met Lys His Leu Asp His Gln Ala Ala Ala Leu Thr Lys Asn Gly Gly	342
90 95 100	
AAG TTT GAG AAG CGG GTG GAC AAT GTG ACA CCT GGG ATC ACC TTG GCC Lys Phe Glu Lys Arg Val Asp Asn Val Thr Pro Gly Ile Thr Leu Ala	390
105 110 115	
ACT AGG GGA CTG TCC CCA TCT GCC CTG ACA AAG CCT GAA TCC GCC ACA Thr Arg Gly Leu Ser Pro Ser Ala Leu Thr Lys Pro Glu Ser Ala Thr	438
120 125 130	
TTG GAA GAC CTT GCT TTG GAA CTG ACT ACT ATT TCC CAG GAG GCC AGG Leu Glu Asp Leu Ala Leu Glu Leu Thr Thr Ile Ser Gln Glu Ala Arg	486
135 140 145	
GGG ACC ATG GGG ACT TCC CAA GAG CCA CCG GCA GCA GTG ACC GGA TCA Gly Thr Met Gly Thr Ser Gln Glu Pro Pro Ala Ala Val Thr Gly Ser	534
150 155 160 165	
TCT CTC TCA ACT TCC GAG GCA CAG GAT GCA GGG CTT ACG GCT AAG CCT Ser Leu Ser Thr Ser Glu Ala Gln Asp Ala Gly Leu Thr Ala Lys Pro	582
170 175 180	
CAG AGC ATT GGA AGT TTT GAG GCG GCT GAC ATC TCC ACC ACC GTT TGG Gln Ser Ile Gly Ser Phe Glu Ala Ala Asp Ile Ser Thr Thr Val Trp	630
185 190 195	
CCG AGT CCT GCT GTC TAC CAA TCT GGA TCT AGC TCC TGG GCT GAG GAA Pro Ser Pro Ala Val Tyr Gln Ser Gly Ser Ser Ser Trp Ala Glu Glu	678
200 205 210	
AAA GCT ACT GAG TCC CCC TCC ACT ACA GCC CCA TCT CCT CAG GTG TCC Lys Ala Thr Glu Ser Pro Ser Thr Thr Ala Pro Ser Pro Gln Val Ser	726
215 220 225	
ACT ACT TCA CCT TCA ACC CCA GAG GAA AAT GTT GGG TCC GAA GGC CAA Thr Thr Ser Pro Ser Thr Pro Glu Glu Asn Val Gly Ser Glu Gly Gln	774
230 235 240 245	
CCC CCA TGG GTC CAG GGA CAG GAC CTC AGT CCA GAG AAG TCT CTA GGG Pro Pro Trp Val Gln Gly Gln Asp Leu Ser Pro Glu Lys Ser Leu Gly	822
250 255 260	

- 74 -

TCT GAG GAG ATA AAC CCA GTT CAT ACT GAT AAT TTC CAG GAG AGG GGG	870
Ser Glu Glu Ile Asn Pro Val His Thr Asp Asn Phe Gln Glu Arg Gly	
265 270 275	
CCT GGC AAC ACA GTC CAC CCC TCA GTG GCT CCC ATC TCC TCT GAA GAG	918
Pro Gly Asn Thr Val His Pro Ser Val Ala Pro Ile Ser Ser Glu Glu	
280 285 290	
ACC CCC AGC CCA GAG CTG GTG GCC TCG GGC AGC CAG GCT CCT AAG ATA	966
Thr Pro Ser Pro Glu Leu Val Ala Ser Gly Ser Gln Ala Pro Lys Ile	
295 300 305	
GAG GAA CCC ATC CAT GCC ACT GCA GAT CCC CAG AAA CTG AGT GTG CTT	1014
Glu Glu Pro Ile His Ala Thr Ala Asp Pro Gln Lys Leu Ser Val Leu	
310 315 320 325	
ATC ACT CCT GTC CCC GAC ACC CAG GCA GCC ACA AGG AGG CAG GCA GTG	1062
Ile Thr Pro Val Pro Asp Thr Gln Ala Ala Thr Arg Arg Gln Ala Val	
330 335 340	
GGG CTA CTG GCT TTC CTT GGT CTT CTT TTC TGC CTA GGG GTG GCC ATG	1110
Gly Leu Leu Ala Phe Leu Gly Leu Leu Phe Cys Leu Gly Val Ala Met	
345 350 355	
TTT GCT TAC CAG AGC CTT CAG GGC TGT CCC CGC AAA ATG GCG GGG GAA	1158
Phe Ala Tyr Gln Ser Leu Gln Gly Cys Pro Arg Lys Met Ala Gly Glu	
360 365 370	
ATG GTA GAA GGC CTC CGC TAC GTC CCC CGT AGC TGT GGC AGT AAC TCA	1206
Met Val Glu Gly Leu Arg Tyr Val Pro Arg Ser Cys Gly Ser Asn Ser	
375 380 385	
TAC GTC CTG GTG CCA GTG TGAGCTGCTT GCCTGCCTGC CTGTGTCCAG AGTGTGAT	1262
Tyr Val Leu Val Pro Val	
390 395	
TCGGACAGCT GTCTGGGGAC CCCCCCCAT CCTCATACCC ACCTTCATCC ACGCTGGGGA	1322
AATGGGAATG GAGAAGCTGG ACCCTCCAGG GGCTGTGGGC TCCATCCAAT CCCCCCTCCC	1382
CCGAGGGGTG GCCCCGGAGG CCACCCTAGA CCACTATTCA CTTATCAGAG ACAGAGCAGG	1442
TGACCTTCCA GCTCCTCTAT ATTTGAAAGA ATCCTCTGCT GCTGGCTGGT TAGAGGGGCC	1502
CTTGACACCC CAACTCCAGT GAACAATTAT TTATTGGATT CCCAGCCCCT GCGACGACAC	1562
CTGTTTCCCG CGCGCACCGT GGTCCGCCCA TATCACAAGC AGCAGGCCAG GCCTATCTGC	1622
CTGTCCCCCT GACCTCCTTG TGTCTCCTGG CTTTGCTGCA GTCGCCAGCC CTTCTCCTCC	1682
CCGCCAGCT GCGGTGCTAT CTGCCCTATG TCTCCCTCTA TCCCCTGTAC AGAGCGCACC	1742
ACCATCACCA TCAAAAAAAAA AAAAAAAAAA AAGGGCGGCC GC	1784

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 395 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

- 75 -

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Ala Pro Ser Pro Leu Ala Trp Leu Leu Arg Leu Ala Ala Phe Phe
 1 5 10 15
 His Leu Cys Thr Leu Leu Pro Gly Gln His Leu Gly Met Thr Lys Cys
 20 25 30
 Glu Ile Met Cys Asp Lys Met Thr Ser Arg Ile Pro Val Ala Leu Leu
 35 40 45
 Ile Arg Tyr Gln Leu Asn Gln Glu Ser Cys Gly Lys Arg Ala Ile Val
 50 55 60
 Leu Glu Thr Thr Gln His Arg Arg Phe Cys Ala Asp Pro Lys Glu Lys
 65 70 75 80
 Trp Val Gln Asp Ala Met Lys His Leu Asp His Gln Ala Ala Ala Leu
 85 90 95
 Thr Lys Asn Gly Gly Lys Phe Glu Lys Arg Val Asp Asn Val Thr Pro
 100 105 110
 Gly Ile Thr Leu Ala Thr Arg Gly Leu Ser Pro Ser Ala Leu Thr Lys
 115 120 125
 Pro Glu Ser Ala Thr Leu Glu Asp Leu Ala Leu Glu Leu Thr Thr Ile
 130 135 140
 Ser Gln Glu Ala Arg Gly Thr Met Gly Thr Ser Gln Glu Pro Pro Ala
 145 150 155 160
 Ala Val Thr Gly Ser Ser Leu Ser Thr Ser Glu Ala Gln Asp Ala Gly
 165 170 175
 Leu Thr Ala Lys Pro Gln Ser Ile Gly Ser Phe Glu Ala Ala Asp Ile
 180 185 190
 Ser Thr Thr Val Trp Pro Ser Pro Ala Val Tyr Gln Ser Gly Ser Ser
 195 200 205
 Ser Trp Ala Glu Glu Lys Ala Thr Glu Ser Pro Ser Thr Thr Ala Pro
 210 215 220
 Ser Pro Gln Val Ser Thr Thr Ser Pro Ser Thr Pro Glu Glu Asn Val
 225 230 235 240
 Gly Ser Glu Gly Gln Pro Pro Trp Val Gln Gly Gln Asp Leu Ser Pro
 245 250 255
 Glu Lys Ser Leu Gly Ser Glu Glu Ile Asn Pro Val His Thr Asp Asn
 260 265 270
 Phe Gln Glu Arg Gly Pro Gly Asn Thr Val His Pro Ser Val Ala Pro
 275 280 285
 Ile Ser Ser Glu Glu Thr Pro Ser Pro Glu Leu Val Ala Ser Gly Ser
 290 295 300
 Gln Ala Pro Lys Ile Glu Glu Pro Ile His Ala Thr Ala Asp Pro Gln
 305 310 315 320

- 76 -

Lys	Leu	Ser	Val	Leu	Ile	Thr	Pro	Val	Pro	Asp	Thr	Gln	Ala	Ala	Thr
				325					330					335	
Arg	Arg	Gln	Ala	Val	Gly	Leu	Leu	Ala	Phe	Leu	Gly	Leu	Leu	Phe	Cys
			340					345					350		
Leu	Gly	Val	Ala	Met	Phe	Ala	Tyr	Gln	Ser	Leu	Gln	Gly	Cys	Pro	Arg
		355					360					365			
Lys	Met	Ala	Gly	Glu	Met	Val	Glu	Gly	Leu	Arg	Tyr	Val	Pro	Arg	Ser
	370					375					380				
Cys	Gly	Ser	Asn	Ser	Tyr	Val	Leu	Val	Pro	Val					
385					390					395					

(2) INFORMATION FOR SEQ ID NO:3:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1654 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 86...1276

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AAGCTTGGCA	CGAGGGCACT	GAGCTCTGCC	GCCTGGCTCT	AGCCGCCTGC	CTGGCCCCCG	60										
CCGGGACTCT	TGCCCCACCT	CAGCC	ATG	GCT	CCG	ATA	TCT	CTG	TCG	TGG	CTG	112				
			Met	Ala	Pro	Ile	Ser	Leu	Ser	Trp	Leu					
			1				5									
CTC	CGC	TTG	GCC	ACC	TTC	TGC	CAT	CTG	ACT	GTC	CTG	CTG	GCT	GGA	CAG	160
Leu	Arg	Leu	Ala	Thr	Phe	Cys	His	Leu	Thr	Val	Leu	Leu	Ala	Gly	Gln	
10					15					20					25	
CAC	CAC	GGT	GTG	ACG	AAA	TGC	AAC	ATC	ACG	TGC	AGC	AAG	ATG	ACA	TCA	208
His	His	Gly	Val	Thr	Lys	Cys	Asn	Ile	Thr	Cys	Ser	Lys	Met	Thr	Ser	
				30					35					40		
AAG	ATA	CCT	GTA	GCT	TTG	CTC	ATC	CAC	TAT	CAA	CAG	AAC	CAG	GCA	TCA	256
Lys	Ile	Pro	Val	Ala	Leu	Leu	Ile	His	Tyr	Gln	Gln	Asn	Gln	Ala	Ser	
			45					50					55			
TGC	GGC	AAA	CGC	GCA	ATC	ATC	TTG	GAG	ACG	AGA	CAG	CAC	AGG	CTG	TTC	304
Cys	Gly	Lys	Arg	Ala	Ile	Ile	Leu	Glu	Thr	Arg	Gln	His	Arg	Leu	Phe	
		60					65					70				
TGT	GCC	GAC	CCG	AAG	GAG	CAA	TGG	GTC	AAG	GAC	GCG	ATG	CAG	CAT	CTG	352
Cys	Ala	Asp	Pro	Lys	Glu	Gln	Trp	Val	Lys	Asp	Ala	Met	Gln	His	Leu	
	75					80					85					
GAC	CGC	CAG	GCT	GCT	GCC	CTA	ACT	CGA	AAT	GGC	GGC	ACC	TTC	GAG	AAG	400
Asp	Arg	Gln	Ala	Ala	Ala	Leu	Thr	Arg	Asn	Gly	Gly	Thr	Phe	Glu	Lys	
90					95					100					105	
CAG	ATC	GGC	GAG	GTG	AAG	CCC	AGG	ACC	ACC	CCT	GCC	GCC	GGG	GGA	ATG	448
Gln	Ile	Gly	Glu	Val	Lys	Pro	Arg	Thr	Thr	Pro	Ala	Ala	Gly	Gly	Met	
				110					115					120		

- 77 -

GAC Asp	GAG Glu	TCT Ser	GTG Val	GTC Val	CTG Leu	GAG Glu	CCC Pro	GAA Glu	GCC Ala	ACA Thr	GGC Gly	GAA Glu	AGC Ser	AGT Ser	AGC Ser	496
			125					130					135			
CTG Leu	GAG Glu	CCG Pro	ACT Thr	CCT Pro	TCT Ser	TCC Ser	CAG Gln	GAA Glu	GCA Ala	CAG Gln	AGG Arg	GCC Ala	CTG Leu	GGG Gly	ACC Thr	544
		140					145					150				
TCC Ser	CCA Pro	GAG Glu	CTG Leu	CCG Pro	ACG Thr	GGC Gly	GTG Val	ACT Thr	GGT Gly	TCC Ser	TCA Ser	GGG Gly	ACC Thr	AGG Arg	CTC Leu	592
		155				160					165					
CCC Pro	CCG Pro	ACG Thr	CCA Pro	AAG Lys	GCT Ala	CAG Gln	GAT Asp	GGA Gly	GGG Gly	CCT Pro	GTG Val	GGC Gly	ACG Thr	GAG Glu	CTT Leu	640
170					175					180					185	
TTC Phe	CGA Arg	GTG Val	CCT Pro	CCC Pro	GTC Val	TCC Ser	ACT Thr	GCC Ala	GCC Ala	ACG Thr	TGG Trp	CAG Gln	AGT Ser	TCT Ser	GCT Ala	688
				190					195						200	
CCC Pro	CAC His	CAA Gln	CCT Pro	GGG Gly	CCC Pro	AGC Ser	CTC Leu	TGG Trp	GCT Ala	GAG Glu	GCA Ala	AAG Lys	ACC Thr	TCT Ser	GAG Glu	736
			205					210					215			
GCC Ala	CCG Pro	TCC Ser	ACC Thr	CAG Gln	GAC Asp	CCC Pro	TCC Ser	ACC Thr	CAG Gln	GCC Ala	TCC Ser	ACT Thr	GCG Ala	TCC Ser	TCC Ser	784
		220					225					230				
CCA Pro	GCC Ala	CCA Pro	GAG Glu	GAG Glu	AAT Asn	GCT Ala	CCG Pro	TCT Ser	GAA Glu	GGC Gly	CAG Gln	CGT Arg	GTG Val	TGG Trp	GGT Gly	832
		235				240					245					
CAG Gln	GGA Gly	CAG Gln	AGC Ser	CCC Pro	AGG Arg	CCA Pro	GAG Glu	AAC Asn	TCT Ser	CTG Leu	GAG Glu	CGG Arg	GAG Glu	GAG Glu	ATG Met	880
250					255					260					265	
GGT Gly	CCC Pro	GTG Val	CCA Pro	GCG Ala	CAC His	ACG Thr	GAT Asp	GCC Ala	TTC Phe	CAG Gln	GAC Asp	TGG Trp	GGG Gly	CCT Pro	GGC Gly	928
				270					275					280		
AGC Ser	ATG Met	GCC Ala	CAC His	GTC Val	TCT Ser	GTG Val	GTC Val	CCT Pro	GTC Val	TCC Ser	TCA Ser	GAA Glu	GGG Gly	ACC Thr	CCC Pro	976
			285					290					295			
AGC Ser	AGG Arg	GAG Glu	CCA Pro	GTG Val	GCT Ala	TCA Ser	GGC Gly	AGC Ser	TGG Trp	ACC Thr	CCT Pro	AAG Lys	GCT Ala	GAG Glu	GAA Glu	1024
		300					305					310				
CCC Pro	ATC Ile	CAT His	GCC Ala	ACC Thr	ATG Met	GAC Asp	CCC Pro	CAG Gln	AGG Arg	CTG Leu	GGC Gly	GTC Val	CTT Leu	ATC Ile	ACT Thr	1072
	315					320					325					
CCT Pro	GTC Val	CCT Pro	GAC Asp	GCC Ala	CAG Gln	GCT Ala	GCC Ala	ACC Thr	CGG Arg	AGG Arg	CAG Gln	GCG Ala	GTG Val	GGG Gly	CTG Leu	1120
330					335				340						345	
CTG Leu	GCC Ala	TTC Phe	CTT Leu	GGC Gly	CTC Leu	CTC Leu	TTC Phe	TGC Cys	CTG Leu	GGG Gly	GTG Val	GCC Ala	ATG Met	TTC Phe	ACC Thr	1168
				350				355						360		
TAC Tyr	CAG Gln	AGC Ser	CTC Leu	CAG Gln	GGC Gly	TGC Cys	CCT Pro	CGA Arg	AAG Lys	ATG Met	GCA Ala	GGA Gly	GAG Glu	ATG Met	GCG Ala	1216
			365					370					375			

- 78 -

GAG GGC CTT CGC TAC ATC CCC CGG AGC TGT GGT AGT AAT TCA TAT GTC 1264
 Glu Gly Leu Arg Tyr Ile Pro Arg Ser Cys Gly Ser Asn Ser Tyr Val
 380 385 390

CTG GTG CCC GTG T GAACTCCTCT GGCCTGTGTC TAGTTGTTTG ATTCAGACAG 1317
 Leu Val Pro Val
 395

CTGCCTGGGA TCCCTCATCC TCATACCCAC CCCCACCCAA GGGCCTGGCC TGAGCTGGGA 1377

TGATTGGAGG GGGGAGGTGG GATCCTCCAG GTGCACAAGC TCCAAGCTCC CAGGCATTCC 1437

CCAGGAGGCC AGCCTTGACC ATTCTCCACC TTCCAGGGAC AGAGGGGGTG GCCTCCCAAC 1497

TCACCCCAGC CCCAAACTC TCCTCTGCTG CTGGCTGGTT AGAGGTTCCC TTTGACGCCA 1557

TCCAGCCCCC AATGAACAAT TATTTATTAA ATGCCCAGCC CTTTCTGAAA AAAAAAAAAA 1617

AAAAAAAAAA AAAAAAAAAA AAAATTCCTG CGGCCCGC 1654

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 397 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ala Pro Ile Ser Leu Ser Trp Leu Leu Arg Leu Ala Thr Phe Cys
 1 5 10 15

His Leu Thr Val Leu Leu Ala Gly Gln His His Gly Val Thr Lys Cys
 20 25 30

Asn Ile Thr Cys Ser Lys Met Thr Ser Lys Ile Pro Val Ala Leu Leu
 35 40 45

Ile His Tyr Gln Gln Asn Gln Ala Ser Cys Gly Lys Arg Ala Ile Ile
 50 55 60

Leu Glu Thr Arg Gln His Arg Leu Phe Cys Ala Asp Pro Lys Glu Gln
 65 70 75 80

Trp Val Lys Asp Ala Met Gln His Leu Asp Arg Gln Ala Ala Ala Leu
 85 90 95

Thr Arg Asn Gly Gly Thr Phe Glu Lys Gln Ile Gly Glu Val Lys Pro
 100 105 110

Arg Thr Thr Pro Ala Ala Gly Gly Met Asp Glu Ser Val Val Leu Glu
 115 120 125

Pro Glu Ala Thr Gly Glu Ser Ser Ser Leu Glu Pro Thr Pro Ser Ser
 130 135 140

Gln Glu Ala Gln Arg Ala Leu Gly Thr Ser Pro Glu Leu Pro Thr Gly
 145 150 155 160

- 79 -

Val	Thr	Gly	Ser	Ser	Gly	Thr	Arg	Leu	Pro	Pro	Thr	Pro	Lys	Ala	Gln	165	170	175
Asp	Gly	Gly	Pro	Val	Gly	Thr	Glu	Leu	Phe	Arg	Val	Pro	Pro	Val	Ser	180	185	190
Thr	Ala	Ala	Thr	Trp	Gln	Ser	Ser	Ala	Pro	His	Gln	Pro	Gly	Pro	Ser	195	200	205
Leu	Trp	Ala	Glu	Ala	Lys	Thr	Ser	Glu	Ala	Pro	Ser	Thr	Gln	Asp	Pro	210	215	220
Ser	Thr	Gln	Ala	Ser	Thr	Ala	Ser	Ser	Pro	Ala	Pro	Glu	Glu	Asn	Ala	225	230	235
Pro	Ser	Glu	Gly	Gln	Arg	Val	Trp	Gly	Gln	Gly	Gln	Ser	Pro	Arg	Pro	245	250	255
Glu	Asn	Ser	Leu	Glu	Arg	Glu	Glu	Met	Gly	Pro	Val	Pro	Ala	His	Thr	260	265	270
Asp	Ala	Phe	Gln	Asp	Trp	Gly	Pro	Gly	Ser	Met	Ala	His	Val	Ser	Val	275	280	285
Val	Pro	Val	Ser	Ser	Glu	Gly	Thr	Pro	Ser	Arg	Glu	Pro	Val	Ala	Ser	290	295	300
Gly	Ser	Trp	Thr	Pro	Lys	Ala	Glu	Glu	Pro	Ile	His	Ala	Thr	Met	Asp	305	310	315
Pro	Gln	Arg	Leu	Gly	Val	Leu	Ile	Thr	Pro	Val	Pro	Asp	Ala	Gln	Ala	325	330	335
Ala	Thr	Arg	Arg	Gln	Ala	Val	Gly	Leu	Leu	Ala	Phe	Leu	Gly	Leu	Leu	340	345	350
Phe	Cys	Leu	Gly	Val	Ala	Met	Phe	Thr	Tyr	Gln	Ser	Leu	Gln	Gly	Cys	355	360	365
Pro	Arg	Lys	Met	Ala	Gly	Glu	Met	Ala	Glu	Gly	Leu	Arg	Tyr	Ile	Pro	370	375	380
Arg	Ser	Cys	Gly	Ser	Asn	Ser	Tyr	Val	Leu	Val	Pro	Val				385	390	395

(2) INFORMATION FOR SEQ ID NO:5:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 15 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TATCGGAGCC ATGGC 15

(2) INFORMATION FOR SEQ ID NO:6:

- 80 -

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGGAAAGAAT TCCTGCCGGG TCAGCACCTC GGCATG 36

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 39 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GGGAAACTCG AGTCATTCTC AAAGTTGCCA CCATTTTTA 39

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CACAGTCCAC CCCTCAG 17

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GCTCTGGTAA GCAAACATGG 20

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TGTGAAGTCC TCTGGCCTGT 20

- 81 -

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAAGGGGCTG GGCATTTAAT 20

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GGGAAAGAAT TCATGGCTCC CTCGCCGCTC GCGTCC 36

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 60 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GGGAAACTCG AGTCATTTAT CATCATCATC TTTATAATCC ACTGGCACCA GGACGTATGA 60

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 8 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Asp Tyr Lys Asp Asp Asp Asp Lys
1 5

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

- 82 -

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Leu	Pro	Gly	Gln	His	Leu	Gly	Met	Thr	Lys	Cys	Glu	Ile	Met
1				5					10				

- 83 -

What is claimed is:

1. A substantially pure neurotactin polypeptide.
2. The substantially pure polypeptide of claim 1, wherein said polypeptide is soluble.
- 5 3. The polypeptide of claim 2, wherein said polypeptide comprises the chemokine-like domain of neurotactin.
4. The polypeptide of claim 3, wherein said polypeptide comprises the extracellular domain of
10 neurotactin.
5. The polypeptide of claim 1, wherein said polypeptide is at least 80% identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4.
- 15 6. The polypeptide of claim 5, wherein said polypeptide is at least 90% identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4.
7. The polypeptide of claim 6, wherein said
20 polypeptide has an amino acid sequence identical to the amino acid sequence from amino acid 22 to amino acid 92 in SEQ ID NO:4.
8. The polypeptide of claim 1, wherein said polypeptide is at least 80% identical to the amino acid
25 sequence from amino acid 22 to amino acid 397 in SEQ ID NO:4.

- 84 -

9. The polypeptide of claim 8, wherein said polypeptide is at least 90% identical to the amino acid sequence from amino acid 22 to amino acid 397 in SEQ ID NO:4.

5 10. The polypeptide of claim 9, wherein said polypeptide has an amino acid sequence identical to the amino acid sequence from amino acid 22 to amino acid 397 in SEQ ID NO:4.

10 11. A substantially pure polypeptide comprising a first portion and a second portion, said first portion comprising a neurotactin polypeptide and said second portion comprising an immunoglobulin constant region.

15 12. A substantially pure polypeptide comprising a first portion and a second portion, said first portion comprising a neurotactin polypeptide and said second portion comprising a detectable marker.

20 13. The polypeptide of claim 12, said detectable marker being selected from the group consisting of β -lactamase, chloramphenicol acetyltransferase (CAT), adenosine deaminase (ADA), aminoglycoside phosphotransferase (neo^r , G418^r), dihydrofolate reductase (DHFR), hygromycin-B-phosphotransferase (HPH), thymidine kinase (TK), β -galactosidase, and xanthine guanine phosphoribosyl-transferase (XGPRT).

25 14. A recombinant nucleic acid encoding a neurotactin polypeptide.

15. The nucleic acid of claim 14, wherein said nucleic acid encodes a soluble neurotactin polypeptide.

- 85 -

16. The nucleic acid of claim 14, wherein said nucleic acid encodes the chemokine-like domain of neurotactin.

17. The nucleic acid of claim 14, wherein said
5 nucleic acid encodes the extracellular domain of neurotactin.

18. A nucleic acid encoding a hybrid polypeptide, said hybrid polypeptide comprising a first portion and a second portion, said first portion comprising a
10 neurotactin polypeptide and said second portion comprising an immunoglobulin constant region.

19. A cell comprising the recombinant nucleic acid of claim 14.

20. A vector comprising the nucleic acid of
15 claim 14.

21. The vector of claim 20, said vector being an expression vector.

22. The vector of claim 20, further comprising a regulatory element.

20 23. The vector of claim 22, wherein the regulatory element is selected from the group consisting of the cytomegalovirus hCMV immediate early gene, the early promoter of SV40 adenovirus, the late promoter of SV40 adenovirus, the lac system, the trp system, the TAC
25 system, the TRC system, the major operator and promoter regions of phage lambda, the control regions of fd coat protein, the promoter for 3-phosphoglycerate kinase, the promoters of acid phosphatase, and the promoters of the

- 86 -

yeast α -mating factors.

24. The vector of claim 22, wherein said regulatory element directs tissue-specific expression.

25. The vector of claim 20, wherein said vector
5 is a plasmid.

26. The vector of claim 20, wherein said vector is a virus.

27. The vector of claim 26, wherein said virus is a retrovirus.

10 28. An antibody that selectively binds to a neurotactin polypeptide.

29. The antibody of claim 28, wherein said antibody is a monoclonal antibody.

30. A pharmaceutical composition comprising the
15 polypeptide of claim 1.

31. A method for determining whether a patient has a disorder associated with abnormal expression of neurotactin, said method comprising quantitating the level of expression of neurotactin in a biological sample
20 obtained from said patient.

32. The method of claim 31, comprising quantitating mRNA encoding neurotactin.

33. The method of claim 31, comprising quantitating neurotactin protein.

- 87 -

34. The method of claim 31, wherein said disorder is an inflammatory disease.

35. A method for treating a patient who has a disorder associated with abnormal expression or activity
5 of neurotactin, said method comprising administering to the patient a compound that modulates the expression or activity of neurotactin.

36. The method of claim 35, wherein said compound inhibits the expression of neurotactin.

10 37. The method of claim 36, wherein said compound comprises a small molecule, an antisense nucleic acid molecule, or a ribozyme.

38. The method of claim 35, wherein said compound inhibits the activity of neurotactin.

15 39. The method of claim 38, wherein said compound comprises an antibody that selectively binds to a neurotactin polypeptide.

40. The method of claim 35, wherein said compound is synthetic.

20 41. The method of claim 35, wherein said compound enhances the expression of neurotactin.

42. The method of claim 35, wherein said compound enhances the activity of neurotactin.

- 88 -

43. A method for inhibiting proliferation of progenitor cells in a patient, said method comprising administering to said patient a substantially pure neurotactin polypeptide capable of inhibiting progenitor
5 cell proliferation.

44. A method of suppressing proliferation of an actively dividing myeloid cell comprising contacting said cell with an effective amount of the polypeptide of claim 1.

10 45. An adjunctive method for use with chemotherapy or radiation therapy in a patient comprising:

(a) administering an effective amount of a polypeptide of claim 1 to the patient, and

15 (b) administering chemotherapy or radiation therapy to the patient in conjunction with said administration of said polypeptide.

46. A method of treating a hyperproliferative myeloid disease in a patient, said method comprising
20 administering to the patient an effective amount of a protein of claim 1.

47. The method of claim 46, wherein said disease is chronic myelogenous leukemia, polycythemia vera, or a hypermegakaryocytopoietic disorder.

25 48. A substantially pure polypeptide that functionally interacts with neurotactin.

49. A nucleic acid encoding the protein of claim 48.

- 89 -

50. A transgenic animal harboring the nucleic acid molecule of claim 14.

51. A method of identifying a compound that modulates expression of neurotactin, said method
5 comprising assessing the expression of neurotactin in the presence and absence of said compound.

52. A method of identifying a compound that modulates the activity of neurotactin, said method comprising assessing the activity of neurotactin in the
10 presence and absence of said compound.

53. An isolated nucleic acid that hybridizes under stringent conditions to the nucleic acid of claim 14.

54. A substantially pure polypeptide encoded by
15 the nucleic acid of claim 53.

55. The nucleic acid of claim 14, said nucleic acid comprising SEQ ID NO:1.

56. The nucleic acid of claim 14, said nucleic acid comprising SEQ ID NO:3.

20 57. The polypeptide encoded by the nucleic acid of claim 55.

58. The polypeptide encoded by the nucleic acid of claim 56.

GTCGACCCACGCGTCCGGCCGAATTCTCTGCACTTCCGCC																				7
ATG GCT CCC TCG CCG CTC GCG																				60
W	L	L	R	L	A	A	F	F	H	L	C	T	L	L	P	G	Q	H	L	27
TTG	CTG	CTG	CGC	CTG	GCC	GCG	TTC	TTC	CAT	TTG	TGT	ACT	CTG	CTG	CCG	GGT	CAG	CAC	CTC	120
G	M	T	K	C	E	I	M	C	D	K	M	T	S	R	I	P	V	A	L	47
GGC	ATG	ACG	AAA	TGC	GAA	ATC	ATG	TGC	GAC	AAG	ATG	ACC	TCA	CGA	ATC	CCA	GTG	GCT	TTG	180
L	I	R	Y	Q	L	N	Q	E	S	C	G	K	R	A	I	V	L	E	T	67
CTC	ATC	CGC	TAT	CAG	CTA	AAC	CAG	GAG	TCC	TGC	GGC	AAG	CGT	GCC	ATT	GTC	CTG	GAG	ACG	240
T	Q	H	R	R	F	C	A	D	P	K	E	K	W	V	Q	D	A	M	K	87
ACA	CAG	CAC	AGA	CGC	TTC	TGT	GCT	GAC	CCG	AAG	GAG	AAA	TGG	GTC	CAA	GAC	GCC	ATG	AAG	300
H	L	D	H	Q	A	A	A	L	T	K	N	G	G	K	F	E	R	R	V	107
CAT	CTG	GAT	CAC	CAG	GCT	GCT	GCC	CTC	ACT	AAA	AAT	GGT	GGC	AAG	TTT	GAG	AAG	CGG	GTG	360
D	N	V	T	P	G	I	T	L	A	T	R	G	L	S	P	S	A	L	T	127
GAC	AAT	GTG	ACA	CCT	GGG	ATC	ACC	TTG	GCC	ACT	AGG	GGA	CTG	TCC	CCA	TCT	GCC	CTG	ACA	420
K	P	E	S	A	T	L	E	D	L	A	L	E	L	T	T	I	S	Q	E	147
AAG	CCT	GAA	TCC	GCC	ACA	TTG	GAA	GAC	CTT	GCT	TTG	GAA	CTG	ACT	ACT	ATT	TCC	CAG	GAG	480
A	R	G	T	M	G	T	S	Q	E	P	P	A	A	V	T	G	S	S	L	167
GCC	AGG	GGG	ACC	ATG	GGG	ACT	TCC	CAA	GAG	CCA	CCG	GCA	GCA	GTG	ACC	GGA	TCA	TCT	CTC	540
S	T	S	E	A	Q	D	A	G	L	T	A	K	P	Q	S	I	G	S	F	187
TCA	ACT	TCC	GAG	GCA	CAG	GAT	GCA	GGG	CTT	ACG	GCT	AAG	CCT	CAG	AGC	ATT	GGA	AGT	TTT	600
E	A	A	D	I	S	T	T	V	W	P	S	P	A	V	Y	Q	S	G	S	207
GAG	GCG	GCT	GAC	ATC	TCC	ACC	ACC	GTT	TGG	CCG	AGT	CCT	GCT	GTC	TAC	CAA	TCT	GGA	TCT	660
S	S	W	A	E	E	K	A	T	E	S	P	S	T	T	A	P	S	P	Q	227
AGC	TCC	TGG	GCT	GAG	GAA	AAA	GCT	ACT	GAG	TCC	CCC	TCC	ACT	ACA	GCC	CCA	TCT	CCT	CAG	720
V	S	T	T	S	P	S	T	P	E	E	N	V	G	S	E	G	Q	P	P	247
GTG	TCC	ACT	ACT	TCA	CCT	TCA	ACC	CCA	GAG	GAA	AAT	GTT	GGG	TCC	GAA	GGC	CAA	CCC	CCA	780
W	V	Q	G	Q	D	L	S	P	E	K	S	L	G	S	E	E	I	N	P	267
TGG	GTC	CAG	GGA	CAG	GAC	CTC	AGT	CCA	GAG	AAG	TCT	CTA	GGG	TCT	GAG	GAG	ATA	AAC	CCA	840
V	H	T	D	N	F	Q	E	R	G	P	G	N	T	V	H	P	S	V	A	287
GTT	CAT	ACT	GAT	AAT	TTC	CAG	GAG	AGG	GGG	CCT	GGC	AAC	ACA	GTC	CAC	CCC	TCA	GTG	GCT	900
P	I	S	S	E	E	T	P	S	P	E	L	V	A	S	G	S	Q	A	P	307
CCC	ATC	TCC	TCT	GAA	GAG	ACC	CCC	AGC	CCA	GAG	CTG	GTG	GCC	TCG	GGC	AGC	CAG	GCT	CCT	960
K	I	E	E	P	I	H	A	T	A	D	P	Q	K	L	S	V	L</			

CCCTTGACACCCCAACTCCAG. ACAATTATTTATTGGATTCCGAGCCCTGCGACGAC	1560
ACCTGTTTCCCGCGCGCACCGTGGTCCGCCCATATCACAAGCAGCAGGCCAGGCCATCT	1620
GCCTGTCCCCCTGACCTCCTTGTGTCTCCTGGCTTTGCTGCAGTCGCCAGCCCTTCTCCT	1680
CCCCGGCCAGCTGCGGTGCTATCTGCCCTATGTCTCCCTCTATCCCTGTACAGAGCGCA	1740
CCACCATCACCATCAAAAAAAAAAAAAAAAAAAGGGCGGCCGC	1784

AAGCTTGGCAGGAGGGCACTGAGCTCTGCCGCTGG/ "AGCCGCTTGCCTGG 54
 -----M-A-P-S L S W L L R L 12
 CCCCCCGGGGACTCTTGGCCACCCTCAGCC ATG GCT CCG ATA TCT CTG TCG TGG CTG CTC CGC TTG 120
 A T F C H L T V L L A G Q H H G V T K C 32
 GCC ACC TTC TGC CAT CTG ACT GTC CTG CTG GCT GGA CAG CAC CAC GGT GTG ACG AAA TGC 180
 N I T C S K M T S K I P V A L L I H Y Q 52
 AAC ATC ACG TGC AGC AAG ATG ACA TCA AAG ATA CCT GTA GCT TTG CTC ATC CAC TAT CAA 240
 Q N Q A S C G K R A I I L E T R Q H R L 72
 CAG AAC CAG GCA TCA TGC GGC AAA CGC GCA ATC ATC TTG GAG ACG AGA CAG CAC AGG CTG 300
 F C A D P K E Q W V K D A M Q H L D R Q 92
 TTC TGT GCC GAC CCG AAG GAG CAA TGG GTC AAG GAC GCG ATG CAG CAT CTG GAC CGC CAG 360
 A A A L T R N G G T F E K Q I G E V K P 112
 GCT GCT GCC CTA ACT CGA AAT GGC GGC ACC TTC GAG AAG CAG ATC GGC GAG GTG AAG CCC 420
 R T T P A A G G M D E S V V L E P E A T 132
 AGG ACC ACC CCT GCC GCC GGG GGA ATG GAC GAG TCT GTG GTC CTG GAG CCC GAA GCC ACA 480
 G E S S S L E P T P S S Q E A Q R A L G 152
 GGC GAA AGC AGT AGC CTG GAG CCG ACT CCT TCT TCC CAG GAA GCA CAG AGG GCC CTG GGG 540
 T S P E L P T G V T G S S G T R L P P T 172
 ACC TCC CCA GAG CTG CCG ACG GGC GTG ACT GGT TCC TCA GGG ACC AGG CTC CCC CCG ACG 600
 P K A Q D G G P V G T E L F R V P P V S 192
 CCA AAG GCT CAG GAT GGA GGG CCT GTG GGC ACG GAG CTT TTC CGA GTG CCT CCC GTC TCC 660
 T A A T W Q S S A P H Q P G P S L W A E 212
 ACT GCC GCC ACG TGG CAG AGT TCT GCT CCC CAC CAA CCT GGG CCC AGC CTC TGG GCT GAG 720
 A K T S E A P S T Q D P S T Q A S T A S 232
 GCA AAG ACC TCT GAG GCC CCG TCC ACC CAG GAC CCC TCC ACC CAG GCC TCC ACT GCG TCC 780
 S P A P E E N A P S E G Q R V W G Q G Q 252
 TCC CCA GCC CCA GAG GAG AAT GCT CCG TCT GAA GGC CAG CGT GTG TGG GGT CAG GGA CAG 840
 S P R P E N S L E R E E M G P V P A H T 272
 AGC CCC AGG CCA GAG AAC TCT CTG GAG CCG GAG GAG ATG GGT CCC GTG CCA GCG CAC ACG 900
 D A F Q D W G P G S M A H V S V V P V S 292
 GAT GCC TTC CAG GAC TGG GGG CCT GGC AGC ATG GCC CAC GTC TCT GTG GTC CCT GTC TCC 960
 S E G T P S R E P V A S G S W T P K A E 312
 TCA GAA GGG ACC CCC AGC AGG GAG CCA GTG GCT TCA GGC AGC TGG ACC CCT AAG GCT GAG 1020
 E P I H A T M D P Q R L G V L I T P V P 332
 GAA CCC ATC CAT GCC ACC ATG GAC CCC CAG AGG CTG GGC GTC CTT ATC ACT CCT GTC CCT 1080
 D A Q A A T R R Q A V G L L A F L G L L 352
 GAC GCC CAG GCT GCC ACC CCG AGG CAG GCG GTG GGG CTG CTG GCC TTC CTT GGC CTC CTC 1140
 F C L G V A M F T Y Q S L Q G C P R K M 372
 TTC TGC CTG GGG GTG GCC ATG TTC ACC TAC CAG AGC CTC CAG GGC TGC CCT CGA AAG ATG 1200
 A G E M A E G L R Y I P R S C G S N S Y 392
 GCA GGA GAG ATG GCG GAG GGC CTT CGC TAC ATC CCC CCG AGC TGT GGT AGT AAT TCA TAT 1260
 V L V P V 397
 GTC CTG GTG CCC GTG TGA ACTCCTCTGGCCTGTGTCTAGTTGTTTGATTTCAGACAGCTGC 1320
 CTGGGATCCCTCATCTCATACCCACCCCAAGGGCCTGGCCTGAGCTGGGATGAT 1380
 TGGAGGGGGGAGGTGGGATCCTCCAGGTGCACAAGCTCCAAGCTCCCAAGGCAATCCCCAG 1440
 GAGGCCAGCCTTGACCAATCTCCACCTTCCAGGGACAGAGGGGGTGGCCTCCCAACTCAC 1500
 CCCAGCCCCAAACTCTCTCTGCTGCTGGCTGGTTAGAGGTTCCTTTGACGCCATCCC 1560

AGCCCCAATGAACAATTATTTA...AAATGCCCGAGCCCTTCTGAAAAAAAAAAAAAAAA
AAAAAAAAAAAAAAAAAAAAAAAAATTCCTGCCGCCGC

1620

1654

Fig. 3

Primary Structure Comparison of Tango-001 to Other Chemokine Subfamilies

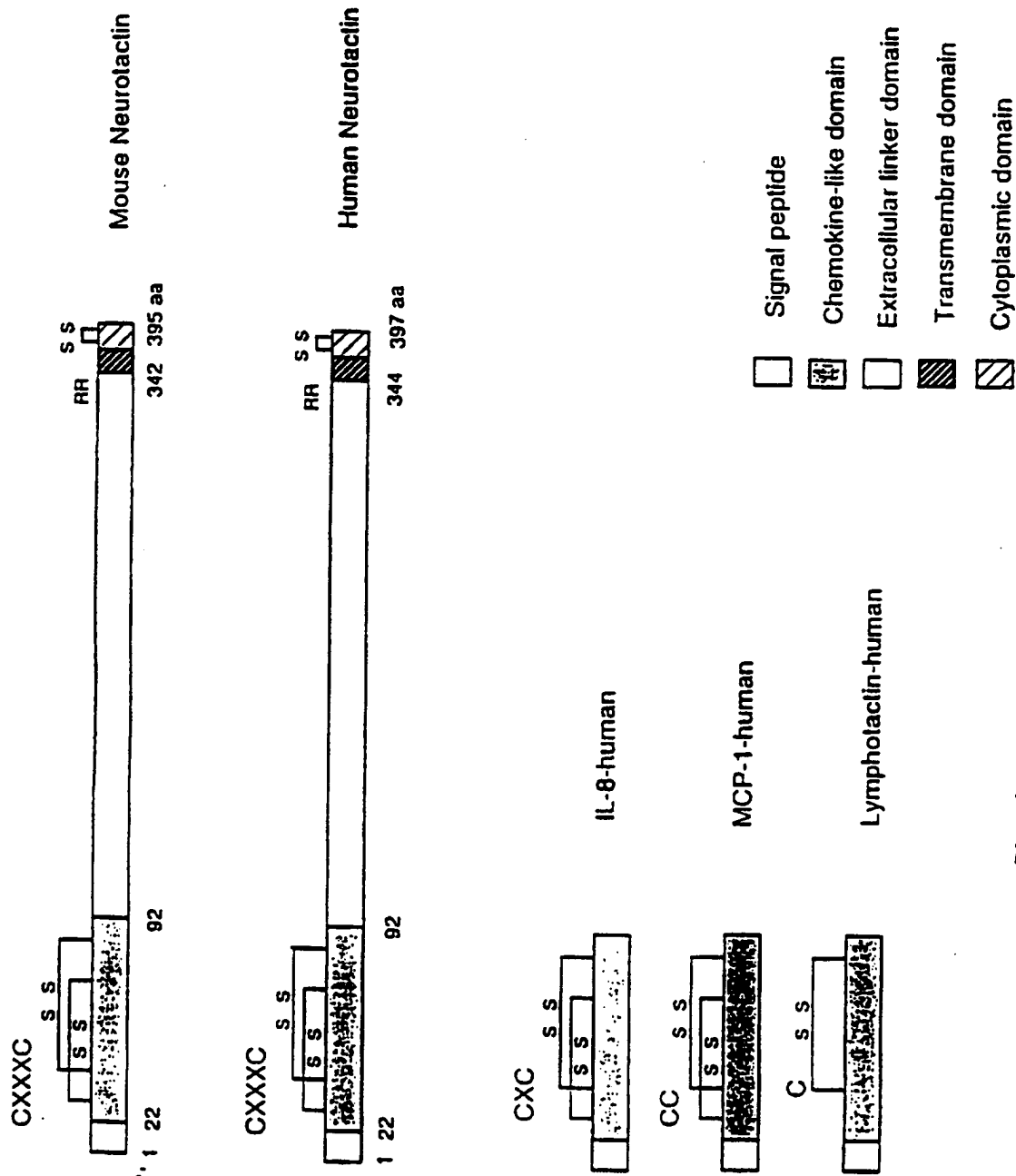
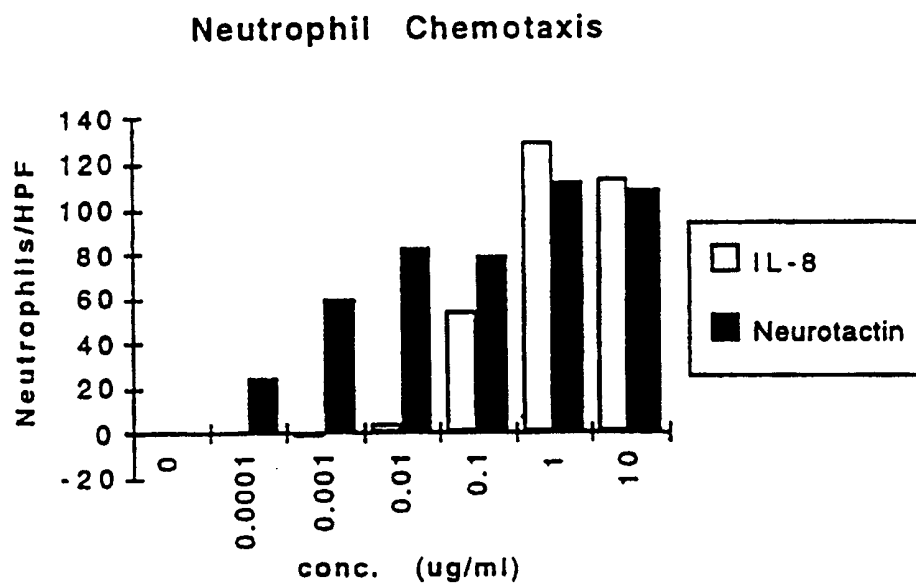


FIG. 4



(HPF, high power field).

Fig. 5

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/07737

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350, 387.3, 388.2; 536/23.1, 23.4, 23.5; 435/7.1, 320.1, 325; 424/130.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
STN medline, embase, biosis, scisearch; APS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A, E	PAN, Y. et al. Neurotactin, a membrane-anchored chemokine upregulated in brain inflammation. Nature. 05 June 1997, Vol. 387, pages 611-617, see entire document.	1-32
X ---- Y	de la ESCALERA, S. et al. Characterization and gene cloning of neurotactin, a Drosophila transmembrane protein related to cholinesterases. EMBO Journal. November 1990, Vol. 9, No. 1, pages 3593-3601, see entire document.	1, 3-4, 14, 16-17, 48 ---- 2, 11-13, 15, 18-25, 28-29, 49, 53
Y	BARTHALAY, Y. et al. Drosophila neurotactin mediates heterophilic cell adhesion. EMBO Journal. November 1990, Vol. 9, No. 11, pages 3603-3609, see entire document.	15-17, 19-27

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

17 JULY 1997

Date of mailing of the international search report

27 AUG 1997

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

F. PIERRE VANDERVEGT

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/07737

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	COCKETT, M.I. et al. The use of engineered E1A genes to transactivate the hCMV-MIE promoter in permanent CHO cell lines. Nucleic Acid Research. 25 January 1991, Vol. 19, No. 2, pages 319-325, especially page 320.	20-24, 26-27
Y	BURKLY, L.C. et al. T-cell tolerance by clonal anergy in transgenic mice with nonlymphoid expression of MHC class II I-E. Nature. 30 November 1989, Vol. 342, pages 564-566, see entire document.	20-22, 24-25, 50

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/07737

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (6):

C07K 16/28, 14/435; C12N 15/00, 15/11, 15/12 5/10; G01N33/53; A61K 39/395

A. CLASSIFICATION OF SUBJECT MATTER:
US CL :

530/350, 387.3, 388.2; 536/23.1, 23.4, 23.5; 435/7.1, 320.1, 325; 424/130.1

THIS PAGE BLANK (USPTO)

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ **BLACK BORDERS**
- ☐ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☐ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☒ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER: _____**

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.

THIS PAGE BLANK (USPTO)